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INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification ⁶: A61K 39/12, C12N 15/14, 15/63	A1	(11) International Publication Number: WO 96/04010 (43) International Publication Date: 15 February 1996 (15.02.96)
(21) International Application Number: PCT/US95/09927 (22) International Filing Date: 4 August 1995 (04.08.95) (30) Priority Data: 08/287,941 5 August 1994 (05.08.94) US (71) Applicant: REGENTS OF THE UNIVERSITY OF MINNESOTA [US/US]; 100 Church Street, S.E., Minneapolis, MN 55455 (US). (72) Inventors: MURTAUGH, Michael, P.; 2957 Northview, Roseville, MN 55113 (US). ELAM, Margaret, R.; 2930 36th Avenue S., Minneapolis, MN 55406 (US). KAKACH, Laura, T.; 106 12th Avenue N., Hopkins, MN 55343 (US). (74) Agent: COLLINS, John, M.; Hovey, Williams, Timmons & Collins, Suite 400, 2405 Grand Boulevard, Kansas City, MO 64108 (US).		(81) Designated States: CA, JP, MX, European patent (AT, BE, CH, DE, DK, ES, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE). Published <i>With international search report.</i>
(54) Title: VR-2332 VIRAL NUCLEOTIDE SEQUENCE AND METHODS OF USE		
(57) Abstract A nucleotide sequence is provided for the VR-2332 virus, which is capable of causing Porcine Reproductive and Respiratory Syndrome. The nucleotide sequence includes protein coding regions that are inserted into recombinant vectors for the host expression of viral proteins according to a variety of vaccination techniques. Diagnostic assays utilize fragmentary sequences or oligonucleotides to selectively identify the VR-2332 nucleic acids by hybridization or PCR amplification reactions that distinguish VR-2332 nucleotide sequences from other PRRS-causative viruses which are immunologically distinct from VR-2332.		

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VR-2332 VIRAL NUCLEOTIDE
SEQUENCE AND METHODS OF USE

5 Sequence Listing

A printed Sequence Listing accompanies this application, and is also submitted with identical contents in the form of a computer-readable ASCII file.

10 Background of the Invention

1. Field of the Invention

The invention pertains to the field of molecular genetics and, in particular, to the use of man-made nucleotides in diagnosing animal diseases or vaccinating animals against disease. More specifically, the preferred nucleotides derive from an immunologically distinct strain of the porcine reproductive and respiratory syndrome ("PRRS") virus, and selectively target this virus in the application of vaccination or diagnostic techniques.

2. Description of the Prior Art

A new viral disease of pigs was detected in North America during 1987, and reported by Hill, *Overview and History of Mystery Swine Disease (Swine Infertility and Respiratory syndrome)*, in Proceedings of the Mystery Swine Disease Committee Meeting, October 6, Denver CO, from the Livestock Conservation Institute of Madison, Wisconsin pp. 29-30 (1990). A disease having substantially identical clinical signs was found in Europe during 1990, as reported by Paton et al., *Blue ear disease of pigs*, 128 Vet Rec. 617 (1991). The clinically observed disease is commonly known by various names including porcine reproductive and respiratory syndrome ("PRRS"), swine infertility and respiratory syndrome ("SIRS"), porcine epidemic abortion and respiratory syndrome ("PEARS"), and mystery swine disease; herein, the term PRRS will suffice to indicate all of these names.

The consequences of this disease included late-term abortions and stillbirths in sows, as well as respiratory insufficiencies in nursery pigs that developed poorly and died easily. Decreases were observed in sow conception rates and litter sizes. Estimates stated that about ten to fifteen percent of pig production were lost annually due to reproductive failure. Early

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clinical signs of the disease included anorexia and mild pyrexia. Other signs included bluish discolorations on the skin of diseased herd animals, with the discolorations being primarily located on the ears, teats, snout, and frontal portions of the neck and shoulders. Necropsy results indicated thickened alveolar septae caused by the presence of macrophages, degenerating cells, and debris in alveolar spaces. These abnormalities indicated the presence of PRRS virus.

The causative viral agent was suspected to be a small, enveloped positive-stranded RNA virus that was recovered primarily from alveolar macrophages of infected swine, as reported by Benfield et al., *Characterization of swine infertility and respiratory syndrome (SIRS) virus (isolate ATCC VR-2332)*, 4 J. Vet. Diagn. Invest. 127-133 (1992); and Wensvoort et al., *Lelystad virus, the cause of porcine epidemic abortion and respiratory syndrome: a review of mystery swine disease research at Lelystad*, 33 Vet. Micro. 185-193 (1992). The isolation technique for the Lelystad ("LV") virus included homogenizing infected swine lung tissue; mixing the homogenate with a physiological saline, e.g., Ringers solution, Hank's balanced salt solution, and Minimum Essential Medium ("MEM") to a 10% weight/volume amount of the homogenate; and filtering the mixture through a series of 0.45, 0.2 and 0.1 micron filters.

The LV virus appeared to be closely related to arteriviruses in morphology, genome organization, transcriptional regulation, and macrophage specificity, according to Plagemann et al., *Lactate dehydrogenase-elevating virus, equine arteritis virus and simian hemorrhagic fever virus: a new group of positive-strand RNA viruses*, 41 Adv. Vir. Res. 99-192 (1992).

The complete nucleotide sequence of the LV strain of the PRRS virus was identified by Meulenbergh et al., *Lelystad virus, the causative agent of porcine epidemic abortion and respiratory syndrome (PEARS), is related to LDV and EAV*, 192 Virology 62-72 (1993). A partial LV sequence was also identified by Conzelmann et al., *Molecular characterization of porcine reproductive and respiratory syndrome virus, a member of the arterivirus group*, 193 Virology 193, 329-339. The positive-strand genome of the LV virus (Sequence ID. Nos. 14-26) included eight open reading frames ("ORFs"), which had some similarity in comparison with the genes of coronaviruses and arteriviruses. Two open reading frames likely coded for the viral RNA

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polymerase. LV ORFs two through six appeared to code for structural proteins associated with viral membranes, and LV ORF 7 was believed to code for a nucleocapsid.

5 The LV viral proteins were expressed from a nested set of RNA transcripts that had overlapping 3' ends. While this expression strategy was shared with the Coronavirus family, the physical properties of the LV virus originally placed it in the Togavirus family. Plagemann et al. (see above) has proposed a new family, the Arteriviridae, to encompass viruses having these dual properties. This family included the PRRS virus, equine arteritis virus ("EAV"), lactate dehydrogenase-elevating virus ("LDV"), and simian hemorrhagic fever virus ("SHFV").

10 A second strain ("VR-2332") of the PRRS virus was isolated as a fourth cell culture passage, as reported by Benfield et al., *Characterization of swine infertility and respiratory syndrome (SIRS) virus (isolate ATCC VR-2332)*, 4 J. Vet. Diagn. Invest. 4, 127-133 (1992). Nevertheless, the viral genome was not sequenced. The VR-2332 isolate was deposited in the American Type Culture Collection, and now has an ATCC catalogue number VR-2332. The VR-2332 virus was characterized as spherical with an average diameter of 62 nm and a 25-30 nm core surrounded by an envelope. Viral particles had a buoyant density of 1.18-1.19 g/ml in cesium chloride and were further purified from filtered tissue homogenates by centrifugation on cesium chloride gradients.

20 The respective VR-2332 and LV virus isolates displayed vast differences in terms of antigenic variation, especially in view of their common morphology and similar clinical signs in swine. A comparison study between 24 field sera and seven viral isolates from Europe and North America failed to distinguish a single common antigen which was able to diagnose infection in a reliable manner for both viruses, as reported by Wensvoort et al., *Antigenic comparison of Lelystad virus and swine infertility and respiratory syndrome (SIRS) virus*, 4 J. Vet. Diagn. Invest. 134-138 (1992). Accordingly, despite the structural and symptomological similarities between the two virus strains, it is unlikely that a single vaccine could be developed from one strain of the virus for purposes of immunizing swine against both strains.

Summary of the Invention

The present invention overcomes the problems that are outlined above by providing man-made nucleotide sequences for the immunologically distinct VR-2332 strain of PRRS virus, as well as vaccines derived from these nucleotides and corresponding methods of vaccination.

Broadly speaking, the present invention includes materials and methods that derive from the VR-2332 form of PRRS pathogen. The materials preferably include VR-2332 virus based nucleic acids and proteins having lengths sufficient to make them unique in comparison with the LV form of PRRS pathogen. The methods involve the use of these materials in diagnostic assays and vaccination procedures.

A particularly preferred material of the present invention includes a purified and isolated nucleic acid coding for a fragmentary portion of the VR-2332 genomic sequence between ORF 2 and ORF 7. These sequences are unique with respect to the LV virus genome, and preferably code for the expression of a polypeptide capable of inducing an anti-VR-2332 PRRS immune response in swine. Despite the similarity in PRRS clinical signs and viral morphology between the VR-2332 and LV viruses, VR-2332 based oligonucleotides can be used as polymerase chain reaction ("PCR") primers for the selective amplification of VR-2332 cDNA. These sequences also include inverse complimentary oligonucleotide sequences derived from the VR-2332 genome. These oligonucleotide sequences are also capable of being used as probes in hybridization studies to selectively identify wild-type VR-2332 cDNA.

Portions of the VR-2332 nucleotide sequence may be recombined with a chimeric vector to place the VR-2332 coding region insert under the control of an appropriate promoter sequence and a termination sequence. This vector may be used for host expression of a protein coded for by the insert. Host expression may be accomplished in either prokaryotic or eukaryotic cells. These vectors may be constructed as recombinant plasmids and injected directly into swine to induce an immune response as the host-swine produces viral proteins. Alternatively, the viral proteins may be produced in cell cultures and injected into swine for immunization purposes.

These nucleotide sequences may also be used in PCR diagnostic assays utilizing primers that selectively amplify either VR-2332-based cDNA or LV-based cDNA. Alternatively, these primer sequences can be used in

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hybridization reactions that indicate the presence of a particular PRRS-causative virus.

Other objects, advantages and salient features of the present invention will become apparent from the following detailed description which, when taken into conjunction with the annexed drawings, discloses a number of embodiments of the present invention.

Brief Description of the Drawings

Figure 1 depicts the positional organization of VR-2332 ORFs 2 through 7 with reference to shaded regions corresponding to cDNA fragments from various clones that were used to determine the nucleotide sequence of the VR-2332 strain of the PRRS virus to yield Sequence ID No. 1;

Fig. 2 depicts the nucleotide and deduced amino acid sequence of VR-2332 ORFs 2 through 7, which correspond to Sequence ID Nos. 1 through 13;

Fig. 3A depicts a comparison between the respective amino acid alignments of ORF 7 for VR-2332 and LV virus according to an IUPAC single letter amino acid code wherein identical residues are represented by capital letters and different residues are represented by lower case letters, and the full three letter amino acid code sequences for these residues are provided in Sequence ID No. 13 (VR-2332) and Sequence ID. No. 26 (LV virus);

Fig. 3B depicts a hydropathy profile for VR-2332 ORF 7, wherein the ordinate represents a hydrophobicity value and the abscissa represents a residue number;

Fig. 3C depicts a hydropathy profile for LV virus ORF 7, which is substantially similar to Fig. 3B;

Fig. 4A depicts a comparison between the respective amino acid alignments of ORF 6 for VR-2332 and LV virus according to an IUPAC single letter amino acid code wherein identical residues are represented by capital letters and different residues are represented by lower case letters, and the full three letter amino acid code sequences for these residues are provided in Sequence ID No. 11 (VR-2332) and Sequence ID. No. 24 (LV virus);

Fig. 4B depicts a hydropathy profile for VR-2332 ORF 6, wherein the ordinate represents a hydrophobicity value and the abscissa represents a residue number;

Fig. 4C depicts a hydropathy profile for LV virus ORF 6, which is substantially similar to Fig. 4B;

5 Fig. 5A depicts a comparison between the respective amino acid alignments of ORF 5 for VR-2332 and LV virus according to an IUPAC single letter amino acid code wherein identical residues are represented by capital letters and different residues are represented by lower case letters, and the full three letter amino acid code sequences for these residues are provided in Sequence ID No. 9 (VR-2332) and Sequence ID. No. 22 (LV virus);

10 Fig. 5B depicts a hydropathy profile for VR-2332 ORF 5, wherein the ordinate represents a hydrophobicity value and the abscissa represents a residue number;

Fig. 5C depicts a hydropathy profile for LV virus ORF 5, which is substantially similar to Fig. 5B;

15 Fig. 6A depicts a comparison between the respective amino acid alignments of ORF 4 for VR-2332 and LV virus according to an IUPAC single letter amino acid code wherein identical residues are represented by capital letters and different residues are represented by lower case letters, and the full three letter amino acid code sequences for these residues are provided in Sequence ID No. 7 (VR-2332) and Sequence ID. No. 20 (LV virus);

20 Fig. 6B depicts a hydropathy profile for VR-2332 ORF 4, wherein the ordinate represents a hydrophobicity value and the abscissa represents a residue number;

Fig. 6C depicts a hydropathy profile for LV virus ORF 4, which is substantially similar to Fig. 6B;

25 Fig. 7A depicts a comparison between the respective amino acid alignments of ORF 3 for VR-2332 and LV virus according to an IUPAC single letter amino acid code wherein identical residues are represented by capital letters and different residues are represented by lower case letters, and the full three letter amino acid code sequences for these residues are provided in Sequence ID No. 5 (VR-2332) and Sequence ID. No. 18 (LV virus);

30 Fig. 7B depicts a hydropathy profile for VR-2332 ORF 3, wherein the ordinate represents a hydrophobicity value and the abscissa represents a residue number;

35 Fig. 7C depicts a hydropathy profile for LV virus ORF 3, which is substantially similar to Fig. 7B;

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Fig. 8A depicts a comparison between the respective amino acid alignments of ORF 2 for VR-2332 and LV virus according to an IUPAC single letter amino acid code wherein identical residues are represented by capital letters and different residues are represented by lower case letters, and the full three letter amino acid code sequences for these residues are provided in Sequence ID No. 3 (VR-2332) and Sequence ID. No. 16 (LV virus);

Fig. 8B depicts a hydropathy profile for VR-2332 ORF 2, wherein the ordinate represents a hydrophobicity value and the abscissa represents a residue number;

Fig. 8C depicts a hydropathy profile for LV virus ORF 2, which is substantially similar to Fig. 8B; and

Fig. 9 depicts a comparison between the respective 3' untranslated regions of VR-2332 and LV virus.

Detailed Description of the Preferred Embodiment

The following non-limiting Examples set forth preferred methods and materials for practicing the present invention.

EXAMPLE 1

GROWTH OF THE VR-2332 VIRUS

A virally pure MA-104 cell line culture of the ATCC VR-2332 virus was obtained for use as viral inoculum, courtesy of Boehringer Ingelheim of Ridgefield, Connecticut.

A culture was prepared for use in propagating the VR-2332 inoculum. The VR-2332 virus was grown in cells from a monkey kidney cell line according to the methods outlined by Gravell et al., 181 *Proc. Soc. Exp. Biol. Med.*, 112-119. Those skilled in the art may alternatively refer to the cell line as the 2621, MA-104 or USU-104 cell line. Uninfected cells were cultured in 50 ml of Eagle's MEM medium (purchased from Life Technologies, Inc., Gaithersburg, MD), which was supplemented with 10% fetal calf serum and 50 μ g/ml gentamicin from Sigma Chemical Co. of St. Louis, MO. Cells were dislodged from the flask surface with trypsin-versene, centrifuged to pellet the cells for separation from the trypsin-versene supernatant, and split 1:4 for subculturing. The cells were maintained in a 5% humidified CO₂ atmosphere

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at 37°C in 75 cm² plastic tissue culture flasks, with media passage at 5-7 day intervals.

The four 50 ml cell cultures were each infected by decanting the growth media and adding the VR-2332 inoculum in 1 ml of growth media having a titer of approximately 10⁵-10⁶ tissue culture infective doses (TCID₅₀). The resultant mixture was incubated for 30 min, after which time was added 30 ml of fresh MEM media containing 4% fetal calf serum. The infected cells were incubated under CO₂ as described above for 24 or 48 hr, and harvested by decanting the media to leave cells adhered to the flask walls.

EXAMPLE 2

CONSTRUCTION OF A cDNA LIBRARY

The harvested cells from Example 1 were washed with phosphate-buffered saline, and lysed by the addition of 5M guanidine isothiocyanate. Total cellular RNA was extracted according to the protocols described by Chomczynski et al., *Single-step method of RNA isolation by acid guanidinium thiocyanate-phenol-chloroform extraction*, 162 Anal. Biochem. 156-159 (1987). Poly A-containing RNA was selected by oligo dT column chromatography using conventional equipment and procedures from Gibco BRL of Gaithersburg, MD.

A cDNA library was constructed in the lambda unidirectional phage vector, UniZap™XR, using Gigapack® II Gold¹ packaging extract and *E. coli* SURE™ cells, as directed by the kit manufacturer (Stratagene, La Jolla, CA). This procedure is summarized below with reference to materials provided in the commercially available kit.

The poly A-selected RNA obtained from 2 ml of cell lysate was reverse transcribed with Moloney murine leukemia virus reverse transcriptase and a synthetic 50 base oligo dT primer containing a sequence including an Xho I restriction site, as follows:

5'-GAGAGAGAGAGAGAGAGAGAACTAGTCTCGAGTTTTTTTTTTTTTTT
TT-3'.

¹UniZap XR, Gigapack II Gold, and SURE are trademarks of Stratagene Corp. of La Jolla, CA.

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The first strand synthesis reaction also contained 5-methyl dCTP. Second strand synthesis was achieved by utilizing DNA polymerase I and the standard dCTP instead of 5-methyl dCTP. The ends of the double stranded cDNA were made blunt with T4 DNA polymerase, and EcoRI adaptors were added with T4 DNA ligase. The adaptors had the following synthetic nucleotide sequences:

5'-AATTCGGCACGAG-3'

3'-GCCGTGCTC-5'

The resulting cDNA was treated with polynucleotide kinase to phosphorylate the 5' ends, digested to completion with Xho I, and purified on a Sephacryl S-400 column.

The cDNA was ligated to the Uni-ZAP™ XR vector arms with DNA ligase and packaged in the high efficiency packaging extract, Gigapack® II Gold. The resulting packaged infectious phage preparation was plated on the *E. coli* cell line SURE™.

EXAMPLE 3

SCREENING THE cDNA LIBRARY BY PCR

Many unsuccessful attempts were made to screen the cDNA library of Example 2 for purposes of identifying VR-2332 positive plaques by polymerase chain reaction using PCR primer sequences derived from the reported LV virus. Synthetic DNA fragments or primers were produced and labeled with ³²P as an indicator according to conventional protocols. These oligonucleotide primers replicated portions of LV virus ORFs 2, 6 and 7, as were reported by Meulenber et al., *Lelystad virus, the causative agent of porcine epidemic abortion and respiratory syndrome (PEARS), is related to LDV and EAV*, 192 *Virology* 62-72 (1993). No PCR amplified nucleotide products were obtained under a variety of conditions.

The observed total failure in PCR amplification of VR-2332 nucleic acid sequences indicated that the two viruses (LV and VR-2332) have considerable nucleotide sequence differences, which are sufficient to prevent specific PCR amplification of VR-2332 cDNA using LV-derived primers. Therefore, an alternative cloning strategy was devised using LV sequences for hybridization, but not for PCR, to determine the nucleotide sequence corresponding to the structural genes of the VR-2332 strain of the PRRS virus.

EXAMPLE 4**SCREENING THE cDNA LIBRARY BY PLAQUE HYBRIDIZATION**

A PCR-generated nucleotide fragment that replicated cDNA from LV ORF 7 (Sequence ID No. 26 of the LV virus) was ³²P-labeled, and used to probe Northern blots obtained using MA-104 cells infected with the VR-2332 virus. Radiographic bands were obtained from infected cells, but not from uninfected cells. These bands indicated that LV and VR-2332 shared similar sequences which were capable of hybridizing despite the failure of PCR screening in Example 3.

Several fifteen cm agar plates containing a total of about 50,000 plaques were screened from duplicate lifts onto NitroPlus nitrocellulose membranes (Micron Separations Inc., Westboro, MA). Positive plaques that hybridized to the corresponding LV virus probe were identified by their corresponding radiographic bands as determined by exposure to x-ray film. These positive plaques were replated and rescreened for confirmation. Hybridization-positive recombinant Uni-ZAP™ XR phage were subjected to *in vivo* excision as described in the Stratagene instruction manual, in order to obtain plasmid DNA for sequence analysis. A summary of the Stratagene procedure is set forth below.

Recombinant phage were combined with *E. coli* XL1-Blue cells as well as ExAssist helper phage at 37°C for 15 min and, thereafter, cultured in rich media for 2-3 hours with shaking at 37°C. The culture was heated to 70°C for 20 min, and clarified by centrifugation. Supernatant containing rescued phagemid was added to SOLR cells and plated on ampicillin-containing agar plates. These bacterial colonies contained recombinant plasmids.

The resultant clones were amplified in liquid culture. DNA was extracted and further analyzed by EcoRI and XhoI restriction endonuclease digestion (10X excess). The sizes of the VR-2332-specific inserts were estimated by electrophoresis in agarose gels with molecular weight standards. Next, the nucleotide sequence of 23 clones was determined at the 3' end by dideoxynucleotide sequencing using Sequenase, ³⁵S-dATP and Stratagene's synthetic M13 -20 primer:

5'-GTAAACGACGGCCAGT-3'.

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Sequencing products were analyzed on 6% denaturing polyacrylamide gels. Twenty of 23 clones had identical 3' sequences, suggesting these clones were coterminally nested. Six of these 20 clones of various sizes, all containing an identical 3' end, were selected for further DNA sequencing.

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EXAMPLE 5

VR-2332 SEQUENCE ANALYSIS

Nucleotide sequence data were obtained for each of the six selected clones of Example 4 by manual dideoxynucleotide sequencing with Sequenase (US Biochemicals, Cleveland, OH) and automated fluorescence sequencing (Applied Biosystems, Foster City, CA).

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Fig. 1 schematically depicts the native positions of the six clones, i.e., those designated 761, 712, 431, 412, 513, and 416, which were chosen for further sequence analysis. The fragment length scale proceeds from 0 to about 3.5 kb, with a positional reference to Sequence ID No. 1. Clones 431, 412, 513 and 416 were sequenced from their 5' ends to overlap with the sequence generated from the next smaller clone. The gap between the 5' end of clone 416 and the beginning of ORF 2, which was sequenced from both clones 712 and 761, was sequenced from both ends by synthesizing VR-2332-specific primers. Additional sequencing was performed to confirm the sequence on the opposite strand. This strategy produced a sequence of 3358 nucleotides, i.e., Sequence ID. No. 1, on both strands from a combination of six independent clones. Fig. 2 depicts this total sequence, together with its deduced amino acid translation.

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Numerous differences between the LV and VR-2332 viruses occurred throughout the 3' genomic sequences that coded for ORFs 2 through 7, as well as the 3' untranslated region. These differences were due to nucleotide substitutions, base deletions and base additions. The sequence divergence arose, presumably, from error-prone replication, and suggests that the viral replicase has poor fidelity and lacks proofreading activity.

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EXAMPLE 6
AMINO ACID RESIDUE SEQUENCE COMPARISON
AND IMMUNOLOGICAL CROSS-REACTIVITY

5 An initial survey indicated that the deduced proteins from these
six VR-2332 ORFs roughly corresponded to known ORFs 2 through 7 in each
of LV virus, LDV, and EAV. Accordingly, a detailed comparative study was
performed to determine differences between the amino acid residue sequences
of the VR-2332 and the LV virus, as well as the other Arteriviridae including
LDV and EAV. The amino acid sequence comparison was performed using
10 GCG (University of Wisconsin, Madison, WI) and Intelligenetics, Inc. (Mountain
View, CA) software. Sequence ID No. 1 includes the VR-2332 sequence for
the 3'-most 3442 bases of the VR-2332 nucleotide sequence, as well as the 5'-
most 84 bases preceding the start of ORF 2. These 3358 nucleotides encode
the structural proteins of the virus, and include six ORFs with each ORF
15 corresponding to Sequence ID Nos. 2-13. These VR-2332 ORFs have varying
degrees of homologies in comparison with LV ORFs 2-7 as well as other
members of the Arteriviridae family including LV virus, LDV, and EAV. More
specifically, a comparative sequence analysis indicates a degree of amino acid
sequence homology between the VR-2332 virus and the LV virus ranging from
20 55% in ORF 5 to 79% in ORF 6. Table 1 provides the results of this Arteri-
viridae family comparison.

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Table 1

Percent Amino Acid Identity
of VR-2332 with LV, LDV and EAV*

ORF	LV	LDV	EAV
2	63	43	23
3	60	41(31)	39(25)
4	70	39	22
5	55	52	28
6	79	52	27
7	64	56	26

*Homologies were determined using the Needleman-Wunsch algorithm to align sequences and dividing the number of identical amino acids by the total number of amino acids in the smaller ORF. Since ORF 3 of LDV and EAV is significantly smaller than VR-2332 ORF 3, the homology based on division by VR-2332 is also shown in parentheses.

While the VR-2332 ORFs were most like those of LV virus, the comparison of VR-2332 to LDV indicated that VR-2332 has shared an evolutionary history with LDV. VR-2332 shared 55% identity with ORF 5 of LV virus, but had the lowest overall degree of homology with LV. The VR-2332 ORF 5 had the greatest degree of overall homology with respect to its LDV counterpart. VR-2332 ORF 5, which had about 52% identity with LDV ORF 5, was only slightly more similar to LV than it was to LDV. When VR-2332 was compared to LDV, the homologies were higher in ORFs 5, 6, and 7 than in ORFs 2, 3, and 4. Other than providing a basis for explaining the observed antigenic variance between these related viruses, the further significance of these divergences is unclear, in part because the functions of proteins derived from ORFs 2, 3, and 4 are unknown.

These amino acid sequence analyses also demonstrated that, with few exceptions, the sequence differences were widely distributed. The principal differences were located in the signal-sequence coding 5' ends of the ORFs, and ORF 4 in the region of amino acid residues 50-70.

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Both VR-2332 and the LV virus have been identified as different infectious agents that cause the PRRS clinical signs, but have demonstrated very little, if any, immunological cross-reactivity, as reported by Wensvoort et al. (see above). Nevertheless, the deduced amino acid sequence from the 3' end of VR-2332 (Sequence ID Nos. 3, 5, 7, 9, 11, and 13) revealed a genomic organization that is characteristic of the Arteriviridae, i.e., overlapping coding regions in different reading frames of Sequence ID No. 1.

A dot-matrix analysis was performed by utilizing the GCG software to compare the predicted protein structures for ORFs 2-7 of VR-2332 and the LV virus. As will be understood by those skilled in the art, the dot matrix analysis was performed according to a conventional technique by utilizing a sliding window of 21 amino acids with a requirement of 13 identical residues at each location. This analysis demonstrated that all of the ORFs were substantially collinear between VR-2332 and LV, i.e., the respective viral structures were very similar despite extensive amino acid diversity. The nearly collinear nature of the VR-2332 and LV ORFs also indicated that the amino acid residue differences did not arise from genomic rearrangements. Table 2 provides a detailed comparison of the various deduced amino acid residues that correspond to the respective ORFs in VR-2332 and LV virus.

Table 2
Comparison of VR-2332 and LV Virus ORFs 2-7

ORF	Amino Acids		Predicted KD		pI		Glycosylation Sites	
	2332	LV	2332	LV	2332	LV	2332	LV
2	256	249	29.4	28.4	11.0	10.2	2	2
3	254	265	29.0	30.6	8.1	9.4	7*	7
4	178	183	19.5	20.0	7.9	6.1	4	4
5	200	201	22.4	22.4	8.3	8.2	3	2
6	174	173	19.0	18.9	11.3	11.9	1	2
7	123	128	13.5	13.8	10.4	11.2	1*	1

*Not all predicted sites are identical.

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While these studies demonstrated that VR-2332 was more closely related to the LV virus than were other members of the Arteriviridae, the homologies were much lower than expected for two viruses that cause the same disease; i.e., substitutions, deletions and additions occurred throughout the comparative sequences. The predicted proteins had different molecular weights, different isoelectric points, and different predicted glycosylation sites (Table 2).

Although the amino acid homologies were substantially less than expected for viruses that appear to cause an identical disease, the findings were consistent with the striking antigenic diversity reported from serological studies by Wensvoort et al. These studies provided an explanation as to why there is observed little, if any, serological cross-reactivity between naturally occurring VR-2332 and LV antigens. Antigenic differences between VR-2332 and LV virus are due to immunological responses of swine to the dissimilar amino acid sequence regions of the viruses.

EXAMPLE 7

HYDROPATHY PROFILE STUDIES

Other characteristics of the predicted proteins including the hydropathy profiles and percent basic character were compared. The results confirmed that the two viruses (LV and VR-2332) had functions and structures that were significantly more similar than was indicated by the amino acid comparison of Example 6 and immunological cross-reactivity reports.

Comparative hydropathy profiles were created utilizing the EUGENE software package from Daniben Systems Inc. of Cincinnati, Ohio, based upon the deduced amino acid residue sequences for VR-2332 (Sequence ID Nos. 2-13) and LV virus (Sequence ID Nos. 14-26). These profiles indicated that the ORFs of VR-2332 and LV virus correspond structurally despite significant amino acid residue sequence differences. These results are consistent with the observed biological similarities, which contrast with the distinct serological properties between the VR-2332 and LV virus isolates.

The hydropathy profiles compared each corresponding ORF in VR-2332 and the LV virus to indicate that protein structures and protein functions were conserved despite the extensive sequence differences. These

profiles demonstrated highly similar regions of uncharged and charged amino acids, and are accurate predictors of similar functionality in membrane associated proteins of regions that either span or do not span the membrane. Thus, the VR-2332 proteins are similar in structure and function to those of LV virus, but extensive amino acid differences in the viral proteins account for the extensive differences in serological cross-reactivity.

Figs. 3, 3A, 3B, and 3C depict the amino acid sequence alignment and hydropathy profiles for ORF 7 of VR-2332 (Sequence ID No. 13) and LV (Sequence ID No. 26). This ORF is located at the 3' end of the LV genome where the nucleocapsid protein has also been mapped in LDV and EAV, as reported by Godeny et al., *Map location of lactate dehydrogenase-elevating virus (LDV) capsid protein (Vp1) gene*, 177 Virol. 768-771 (1990), and de Vries et al., *Structural proteins of equine arteritis virus*, 66 J. Virol. 6294-6303 (1992). ORF 7 most likely forms the nucleocapsid protein in the PRRS virus. The protein was 64% similar between VR-2332 and LV virus, and VR-2332 ORF 7 was smaller by five amino acids. Nevertheless, the N-terminal half of both proteins encoded by ORF 7 was 26-28% basic and the hydrophobicity profiles were nearly identical. The basic residues presumably facilitate interactions with the RNA genome.

Figs. 4, 4A, 4B, and 4C depict the amino acid sequence alignment and hydropathy profiles for ORF 6 of VR-2332 (Sequence ID No. 11) and LV (Sequence ID No. 24). ORF 6 was the VR-2332 protein that was most similar to its LV virus counterpart, and was the only ORF that coded for an apparent amino terminal signal sequence. The LV and VR-2332 proteins shared 79% identity and one predicted glycosylation site (the LV virus had an additional site not found in VR-2332). Hydropathy profiles of ORF 6 of VR-2332, LV and EAV all showed three highly hydrophobic regions in the N-terminal half of the protein that indicate membrane spanning domains. These regions appear to be a conserved characteristic of all members of the Arteriviridae.

Figs. 5, 5A, 5B, and 5C depict the amino acid sequence alignment and hydropathy profiles for ORF 5 of VR-2332 (Sequence ID No. 9) and LV (Sequence ID No. 22). ORF 5 appears to encode an envelope protein in the Arteriviridae because of its hydropathy profile and putative glycosylation sites. Similarly, according to de Vries et al. (see above) the G_L or ORF 5

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protein for EAV is glycosylated, VR-2332 ORF 5 contains three potential glycosylation sites, two of which are shared with LV. The LV and VR-2332 hydropathy profiles are highly similar although their percent identity (55%) was the lowest of all ORFs. In particular, only seven residues in the amino terminal 40 amino acids are the same, yet the hydropathy profiles are virtually identical. Potential membrane spanning domains between residues 65 and 130 are more pronounced in VR-2332.

Figs. 6, 6A, 6B, and 6C depict the amino acid sequence alignment and hydropathy profiles for ORF 4 of VR-2332 (Sequence ID No. 7) and LV (Sequence ID No. 20). After ORF 6, ORF 4 is the most highly conserved ORF. The carboxyl terminus also is exceptionally hydrophobic in both viruses. Five putative membrane spanning domains are much more distinct in VR-2332 than in LV virus.

Figs. 7, 7A, 7B, and 7C depict the amino acid sequence alignment and hydropathy profiles for ORF 3 of VR-2332 (Sequence ID No. 7) and LV (Sequence ID No. 18). ORF 3 is 60% similar between VR-2332 and LV virus. Nevertheless, ORF 3 is the least similar protein between the two viruses based on hydropathy profiles and by carboxyl terminal deletions of 12 amino acids in VR-2332. As a result of these differences, the corresponding LV protein has a strongly hydrophilic region centered on residue 240, whereas the VR-2332 protein appears amphipathic in this region. The nominal molecular mass of ORF 3 is approximately 30 kD, but it contains seven potential glycosylation sites in each virus, so that its apparent size can be significantly greater.

Figs. 8, 8A, 8B, and 8C depict the amino acid sequence alignment and hydropathy profiles for ORF 2 of VR-2332 (Sequence ID No. 5) and LV (Sequence ID No. 16). ORF 2 was determined to be the largest of the 3' ORFs in VR-2332, and coded for the expression of 256 amino acids. It had a highly basic isoelectric point of 11.0, which was exceeded only by ORF 6, which had a pI of 11.3. The differences in amino acid sequence between VR-2332 and LV virus were distributed throughout the ORF, but the principal effect on the hydropathy profile appeared in the amino terminus.

Fig. 9 VR-2332 depicts an alignment of the 3' untranslated sequence following ORF 7 in VR-2332 and LV virus. This region consisted of 151 nucleotides and a poly A tail of 19 to 20 bases in VR-2332. Similarly, the

LV virus had a noncoding region of 115 bases. Bases 50-171 of the VR-2332 non-coding region of shared a strong homology to bases 13-135 of the LV non-coding region.

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EXAMPLE 8

ISOLATION OF VR-2332 RNA

Viral RNA from infected cell supernatants is isolated for use in reverse transcription and PCR amplification reactions that selectively amplify either the VR-2332 or the LV viral nucleotides as a diagnostic tool for LV or PRRS. Additionally, PCR amplification is used to produce quantities of nucleotides for use in vaccines.

As a diagnostic measure, swine lung tissue homogenates are preferably obtained by selecting tissue samples from alveolar abnormalities that are typical of PRRS; homogenizing these samples; mixing the homogenate with an appropriate physiological saline, e.g., Minimum Essential Medium, to a 10% (w/v) tissue concentration; and filtering the homogenate mixture through a series of filters having 0.45, 0.2 and 0.1 micron openings.

The filtered homogenate is used as inoculum to infect cells of an appropriate cell line, e.g., monkey kidney cells or MA-104. The inoculated culture is incubated until a culture stock is obtained having a high virus titer from about log 5 to log 7.

A first solution is prepared to include 5 M guanidinium isothiocyanate, 50 mM Tris HCl pH 7.5, 25 mM EDTA, 0.5 w/v Sarcosyl, and 1% (v/v) 2-mercaptoethanol. A 10 ml aliquot of this solution is mixed with 100 microliters of 2-mercaptoethanol. A 2 ml portion of the virus stock culture is mixed in a tube with 2 ml of the first solution aliquot, as is 0.4 ml of 2 M sodium acetate, 4 ml phenol, and 1 ml of a chloroform-isoamyl alcohol solution mixed at a ratio of 24 parts of chloroform to 1 part of isoamyl alcohol. The virus-containing mixture is vortexed briefly after the addition of each reagent. The final mixture is vortexed for thirty seconds, chilled on ice for 15 seconds, then centrifuged at 8000 rpm for 20 minutes at 4°C in a JA-20 rotor. The aqueous phase will separate to the top upon centrifugation, and contains the RNA of interest.

The aqueous phase is decanted and transferred to a new tube. About 4 ml of sterile water containing 2% by volume of diethylpyrocarbonate

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before autoclaving, is added to this second tube, as is 4 ml phenol, and 1.6 ml of the 24:1 chloroform-isoamyl alcohol mixture. These ingredients are vortexed, chilled on ice for 15 minutes, centrifuged at 8000 rpm for 20 minutes at 4°C in a JA-20 rotor, and the aqueous phase is again extracted. The resultant aqueous extract is mixed with an equal volume of isopropanol, and chilled on ice for 1 hour to precipitate the RNA.

The precipitated RNA is sedimented by centrifugation at 8000 rpm for 20 minutes at 4°C in a JA-20 rotor. The isopropanol is decanted, and the invisible RNA pellet is dissolved in 0.3 ml of a solution containing 5 M guanidinium isothiocyanate, 50 mM Tris HCl pH 7.5, 25 mM EDTA, 0.5% Sarcosyl, and 1% 2-mercaptoanol, and 0.1% 2-mercaptoethanol. The solution containing the dissolved pellet is transferred to a 1.5 ml microfuge tube, and the RNA is again precipitated with 0.3 ml of isopropanol for 1 hour on ice. The chilled solution is centrifuged at 15,000 rpm in a microfuge for 10 minutes, after which the isopropanol is decanted. The resultant pellet is washed with about 0.5 ml of a solution containing 75% ethanol mixed with 25% water containing 0.2% diethyl pyrocarbonate by volume. After washing, the mixture is vortexed, and centrifuged for 5-10 minutes. The alcohol is decanted, and the RNA pellet is vacuum-dried for about 3 minutes. The pellet is dissolved in 50 ml of water containing 0.2% diethylpyrocarbonate by volume.

EXAMPLE 9

REVERSE TRANSCRIPTION OF RNA TO FORM cDNA

The solution from Example 8 containing RNA and the 0.2% diethylpyrocarbonate water is next subjected to reverse transcription of the RNA to produce complimentary fragments of cDNA. This procedure is preferably conducted by using commercially available kits, such as the RT-PCR kit from Perkin-Elmer. The kits are used according to the manufacturers instructions, which describe the proper use of kit reagents.

By way of example, a master mixture is prepared from named reagents of the RT-PCR kit by mixing 4 ul MgCl₂, 2 ul of 10X buffer, 2 ul dGTP, 2 ul dATP, 2 ul dCTP, 2 ul TTP, 1 ul RNase inhibitor, and 1 ul of reverse transcriptase. A 3 ul aliquot of the RNA and 0.2% diethylpyrocarbonate water mixture is placed into a microfuge tube taking care, if necessary, to dilute the aliquot with 0.2% diethylpyrocarbonate water so as to include no more than 1

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5 µg of total RNA in the tube. The kit contains a mixture of random hexamers, and 1 ul of this mixture is added to the RNA and diethylpyrocarbonate water. The solution then is optionally heated to a temperature from about 65-70°C for 5 to 10 minutes, and placed on ice. The 16 ul of master mix is added to the sample, and incubated at room temperature for about 10 minutes. Thereafter, the tube is incubated in a thermal cycler under the following conditions: 42°C for 15 minutes, 99°C for 5 minutes, and 5°C for 5 minutes. The tube is removed from the thermal cycler and stored at 4°C. The result of this reverse transcriptase reaction contains cDNA, which is subsequently subjected to PCR amplification.

EXAMPLE 10

SELECTIVE PCR AMPLIFICATION OF cDNA

15 In preparation for PCR amplification, a master mixture of the following reagents is prepared. 1 ul of $MgCl_2$, 2 ul of 10X buffer, 0.5 ul of 5' primer, 0.5 ul of 3' primer, 15.875 ul of sterile water, and 0.125 ul of Taq polymerase. The 5' and 3' primers should have a concentration of approximately 10 uM, and are preferably comprised of synthetic nucleotides based upon the sequences listed below in Table 3. A 5 ul aliquot of the reverse transcriptase reaction solution from Example 9 is added to 20 ul of master mixture. The resultant 25 ul combination of master mixture and reverse transcriptase cDNA aliquot is overlain in a tube with 100 ul of mineral oil. The tube is incubated in a thermal cycler under the following conditions: 93°C for 4 minutes for one cycle; 55°C for 30 seconds, 72°C for 45 seconds, and 93°C for 45 seconds, for 30 cycles; and 55°C for 30 seconds, followed by 72°C for 10 minutes for one cycle. After these 32 cycles, the solution is then maintained at 4°C until it is removed from the thermal cycler. The resultant solution, which contains PCR-amplified cDNA, is analyzed on an agarose gel.

25 The preferred agarose gel includes 1.5% agarose mixed with TAE buffer, i.e., 1.5 grams of agarose per 100 ml of buffer. The mixture is melted in a microwave, and 1 ul of 10 mg/ml ethidium bromide solution is added per 100 ml of the gel. The mixture is poured into a casting stand, and allowed to harden for 30-45 minutes. A 5 ul aliquot of the PCR reaction solution is added into a tube, and 2 ul of a UV-sensitive running dye is added to the aliquot. An additional aliquot of 1-2 ul of an appropriate molecular weight marker is also

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added, such as a 100 base ladder from Gibco-BRL. The gel is placed in an electrophoresis chamber and the chamber is filled with a conventional TAE running buffer. Samples are loaded, and run at 80 volts for 1 hour. The electrophoresed PCR products are visualized under UV light. The PCR generated fragments that are visualized under UV light after the agarose gel electrophoresis are subjected to DNA sequencing for unambiguous confirmation of the identity of the viral nucleotide product.

EXAMPLE 11

OLIGONUCLEOTIDE DESIGN FOR SELECTIVE PCR AMPLIFICATION OR HYBRIDIZATION

The 5' and 3' primers that are used in the PCR amplification of Example 10 are preferably constructed, according to conventional protocols or on commercial order, as synthetic nucleotide sequences that replicate regions of interest in the VR-2332 genome. The primer design preferably includes selecting appropriate primers as the entire amino acid-coding sequences of the viral protein, selected ORFs, or, most preferably, coding regions for amino acid sequences representing protein fragments.

The preferred oligonucleotides are selected to include those which specifically target small portions of the VR-2332 coding region, but are incapable of annealing with LV-derived nucleotides. These preferred oligonucleotides are used as primers for PCR amplification techniques to replicate long sequences of cDNA that are selected by the primers for use in vaccines and methods of vaccination. Similarly, the oligonucleotides are also used as probes for subsequent hybridization, cloning, and host expression of protein fragments and nucleotide products for subsequent use in vaccines.

Preferred examples of the cDNA coding regions for expressed protein fragments that are selected for use in producing vaccines include those in which the translated amino acid terminal hydrophobic sequences are removed, as these terminal sequences are usually not present on mature forms of the viral protein. Selected cDNA coding regions can also code for protein fragments in which putative membrane-spanning sequences are removed, as the membrane-spanning sequences likely will not induce immune responses, and this removal generally simplifies the production of immunologically-sensitive proteins by recombinant DNA techniques.

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The sequences listed in Table 3 below represent exemplary primers with positional reference to the accompanying Sequence Listing. All sequences are provided in a 5' to 3' orientation. By way of example, Primer A represents the sequence 5'-GCTGTTAAACAGGGAGTGG-3'. Primer A' is the inverse compliment of the sequence 5'-GTCACCTATTCAATTAGGG-3' (Sequence ID No. 1 positions 3271-3289), i.e., the sequence 5'-CCCTAATTGAATAGGTGAC-3' in which reverse-ordered complimentary nucleotides have been substituted for the sequence at positions 3271-3289.

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Table 3

Primer	Description	Positional Reference		
		Seq. ID	From	To
A	VR-2332 ORF 7 based primer	1	2783	2801
A'	VR-2332 ORF 7 based inverse compliment of the VR-2332 sequence	1	3271	3289
B	VR-2332 ORF 6 based primer	1	2289	2307
B'	VR-2332 ORF 6 based inverse compliment of the VR-2332 sequence	1	2862	2880
C	LV ORF 6 based primer	14	14112	14131
C'	LV ORF 6 based inverse compliment of the LV sequence	14	14551	14570
D	LV ORF 7 based primer	14	14575	14594
D'	LV ORF 7 based inverse compliment of the LV sequence	14	14955	14974
E	VR-2332 ORF 7 based primer *	1	2814	2832
E'	VR-2332 ORF 7 based inverse compliment of the VR-2332 sequence **	1	3273	3291
F	VR-2332 ORF 7 based primer ***	1	2816	2834
F'	VR-2332 ORF 7 based inverse compliment of the VR-2332 sequence ****	1	3181	3198

*A synthetic oligonucleotide may be constructed to include a BamHI restriction site with this sequence, i.e., the additional 5'-GCGGATCC nucleotides, for insertion into Pharmingen's pAcGP67B plasmid vector.

**A synthetic oligonucleotide may be constructed to include an inverse complimentary EcoRI restriction site with this sequence, i.e., the additional 5'-CCGAATTC nucleotides, for insertion into Pharmingen's pAcGP67B plasmid vector.

***A synthetic oligonucleotide may be constructed to include a NdeI restriction site with this sequence, i.e., the additional 5'-GCGCA nucleotides, for insertion into Novagen's pET25b plasmid vector.

****A synthetic oligonucleotide may be constructed to include an inverse complimentary HindIII restriction site with this sequence, i.e., the additional 5'-GCGAAGCT nucleotides, for insertion into Novagen's pET25b plasmid vector.

Primers A and A' of Table 3 will selectively amplify the VR-2332 ORF 7 protein-coding nucleotides in a manner that distinguishes the VR-2332 nucleotides from other viral nucleotide isolates, including LV isolates. Similarly, Primers B and B' will selectively amplify the VR-2332 ORF 6 protein-coding nucleotides in a manner that distinguishes the VR-2332 nucleotides from other viral nucleotide isolates. On the other hand, Primers C and C', will selectively amplify the ORF 6 coding region of LV virus without amplifying VR-2332 ORF 6. Primers D and D' will selectively amplify LV ORF 7 without amplification of VR-2332 ORF 7.

The preferred oligonucleotides of Table 3 are used for diagnosis of the specific PRRS-causative strain or virus through attempted PCR amplification of cDNA or conventional hybridization reactions. By way of example, if the PRRS signs are confirmed clinically in a diseased animal and if the primers that are specific for amplification of the Lelystad virus (e.g., Primers C, C' and D, D') fail to produce cDNA amplification in the PCR reaction, then the absence of LV cDNA would be consistent with a diagnosis of VR-2332 infection. On the other hand, the failure of VR-2332 primers A, A' or B, B' in PCR amplification would be consistent with a diagnosis of LV infection.

In cases where the presence of viral cDNA is confirmed by hybridization to these primer or probe sequences of Table 3, the hybridization occurs in solution with either cDNA or RNA affixed to a solid support such as nitrocellulose or nylon membranes. The recovered hybridized product is detected by conventional radioactive or non-radioactive techniques, which indicate the presence of viral nucleic acid sequence. Those skilled in the art will understand that an elementary list of diagnostic techniques includes dot-blot hybridization, slot-blot hybridization, solution hybridization, southern blot, northern blot, and RNase protection assays.

EXAMPLE 12

CLONING OF VR-2332 PROTEIN CODING SEQUENCES IN HOST EXPRESSION SYSTEMS FOR THE PRODUCTION OF RECOMBINANTLY DERIVED VIRAL PROTEINS

Selected portions of the VR-2332 nucleotide sequence (Sequence ID Nos. 1, 2, 4, 6, 8, 10, and 12) are used to clone an open reading frame, or a plurality of open reading frames, into a commercially available

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plasmid, that is designed for protein expression in a host organism. Examples of commercially available or self-designated systems that are used for the expression of viral proteins in eukaryotic or prokaryotic cells follow.

5 The commercially available eukaryotic baculovirus system from Pharmingen of San Diego, California, which includes the vector pAcGP67B is preferred for use with Primers C and C'. As indicated in Table 3, Primers C and C' may be provided with respective BamHI and EcoRI restriction sites formed of synthetically joined nucleotides for use in linking these primers with the pAcGP67B vector. By this method, the resultant amplified cDNA would
10 incorporate substantially the entire coding region of VR-2332 ORF 7, and would also have a 5'-most BamHI site as well as a 3'-most EcoRI site. These restriction sites are used to place the VR-2332 coding region under the control of the appropriate pAcGP67B promoter and termination sequences for eukaryotic host expression of VR-2332 ORF 7 proteins.

15 Prokaryotic host expression of viral proteins is accomplished in a variety of commercially available host expression systems. The PET system from NovaGen of Madison, Wisconsin is preferred for prokaryotic expression, and includes the vector pET25b. The PET system is preferred for use with Primers D and D', which may be provided with respective NdeI and HindIII
20 restriction sites for use in placing the VR-2332 ORF 7 coding region under the control of appropriate promoter and termination sequences.

The protein corresponding to VR-2332 ORF 7 of Sequence ID Nos. 12 and 13 is expressed by amplifying selected protein coding sequences corresponding to the putative mature protein of ORF 7. This amplification
25 procedure will follow the RT-PCR amplification procedure that is outlined in Examples 8, 9, and 10. The PCR primers are preferably designed to include NdeI and HindIII restriction sites for cloning into the pET25b vector. These sites will result in a protein without a pelB leader or HisTag sequence, which provide alternative options for other expression systems. The mature protein
30 is expressed without a signal peptide sequence by beginning the nucleotide sequence to code for either amino acid number 20 or number 30. The PCR fragments are cloned into the pET25b vector-amplified sequence and used in a host expression system.

35 In selecting protein coding regions other than ORF 7, it is advantageous to delete or truncate certain protein coding regions, e.g., deletion

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of the membrane-spanning C-terminal 17 amino acids from ORF 4 will likely direct antibody responses to biologically relevant portions of the protein.

The recombinant clones are transformed into BL21 cells for induction by isopropyl- β -D-thiogalactopyranoside ("IPTG"). After induction and an appropriate incubation, the expressed recombinant bioprotein is detected on a gel by comparing lysates from induced and uninduced cells. Inclusion body preps are washed with urea or guanidine at a concentration that removes contaminating proteins without solubilizing the ORF 4 protein. Aggregates are resolubilized in urea and refolded in oxidized and reduced glutathione. The resultant soluble, dialyzed protein is further purified by ion-exchange and size exclusion chromatography.

EXAMPLE 13

INDUCTION OF AN IMMUNE RESPONSE IN AN ANIMAL BY INJECTION OF RECOMBINANT VIRAL PROTEINS

The purified proteins from bacterial or eukaryotic expression systems, as produced in Example 12, are injected into animals by conventional immunization routes to elicit immune responses sufficient to immunize the animal against the VR-2332 strains of PRRS virus. The proteins alone, or in combination with a conventional adjuvant, are administered by intramuscular injection, intradermal injection, subcutaneous injection, or otherwise.

As an alternative, live molecularly engineered bacteria or virus that express proteins corresponding to VR-2332 sequences are administered to animals by injection of the expression of VR-2332 proteins in vivo. This in vivo expression of recombinant proteins will also elicit an immune response to the VR-2332 virus.

EXAMPLE 14

THE USE OF VR-2332 DNA TO INDUCE A DIRECT IMMUNE RESPONSE IN AN ANIMAL

VR-2332 based oligonucleotide fragments, which code for ORFs or fragmentary portions of ORFs, are used to generate a direct immune response in an animal. This method generally follows the procedure described in Omer et al., 259 Science 1745-1749 (1993). The DNA is preferably included in plasmid constructs that are grown in bacteria, purified, and injected into

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animals by intramuscular injection, intradermal injection, or by other routes. The injected animal will typically express the cloned protein, and produce a corresponding immune response to the protein that is expressed.

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SEQUENCE LISTING

(1) GENERAL INFORMATION:

- (i) APPLICANT: Murtaugh, Michael P.
- (ii) TITLE OF INVENTION: VR-2332 VIRAL NUCLEOTIDE SEQUENCE AND METHODS OF USE
- (iii) NUMBER OF SEQUENCES: 26
- (iv) CORRESPONDENCE ADDRESS:
 - (A) ADDRESSEE: John M. Collins
 - (B) STREET: 1101 Walnut, Suite 1400
 - (C) CITY: Kansas City
 - (D) STATE: Missouri
 - (E) COUNTRY: USA
 - (F) ZIP: 64106
- (v) COMPUTER READABLE FORM:
 - (A) MEDIUM TYPE: Floppy disk
 - (B) COMPUTER: IBM PC compatible
 - (C) OPERATING SYSTEM: PC-DOS/MS-DOS
 - (D) SOFTWARE: PatentIn Release #1.0, Version #1.25
- (vi) CURRENT APPLICATION DATA:
 - (A) APPLICATION NUMBER:
 - (B) FILING DATE:
 - (C) CLASSIFICATION:
- (viii) ATTORNEY/AGENT INFORMATION:
 - (A) NAME: Collins, John M.
 - (B) REGISTRATION NUMBER: 26122
 - (C) REFERENCE/DOCKET NUMBER: 22907
- (ix) TELECOMMUNICATION INFORMATION:
 - (A) TELEPHONE: (816) 474-9050
 - (B) TELEFAX: (816) 474-9057

(2) INFORMATION FOR SEQ ID NO:1:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 3358 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: unknown
- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO

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(vi) ORIGINAL SOURCE:

- (A) ORGANISM: Arteriviridae (Unclassified)
- (B) STRAIN: VR-2332

(ix) FEATURE:

- (A) NAME/KEY: misc_feature
- (B) LOCATION: 1..768
- (C) IDENTIFICATION METHOD: experimental
- (D) OTHER INFORMATION: /evidence= EXPERIMENTAL
/standard_name= "VR-2332 ORF2"

(ix) FEATURE:

- (A) NAME/KEY: misc_feature
- (B) LOCATION: 624..1385
- (D) OTHER INFORMATION: /standard_name= "VR-2332 ORF 3"

(ix) FEATURE:

- (A) NAME/KEY: misc_feature
- (B) LOCATION: 1169..1701
- (D) OTHER INFORMATION: /standard_name= "VR-2332 ORF 4"

(ix) FEATURE:

- (A) NAME/KEY: misc_feature
- (B) LOCATION: 1716..2315
- (D) OTHER INFORMATION: /standard_name= "VR-2332 ORF 5"

(ix) FEATURE:

- (A) NAME/KEY: misc_feature
- (B) LOCATION: 2303..2824
- (D) OTHER INFORMATION: /standard_name= "VR-2332 ORF 6"

(ix) FEATURE:

- (A) NAME/KEY: misc_feature
- (B) LOCATION: 2817..3185
- (D) OTHER INFORMATION: /standard_name= "VR-2332 ORF 7"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

ATGAAATGGG GTCCATGCAA AGCCTTTTTG ACAAATTGG CCAACTTTT GTGGATGCTT	60
TCACGGAGTT CTTGGTGTCC ATTGTTGATA TCATTATATT TTTGGCCATT TTGTTTGGCT	120
TCACCATCGC CGTTGGCTG GTGGTCTTTT GCATCAGATT GGTTTGCTCC GCGATACTCC	180
GTACGCGCCC TGCCATTCAC TCTGAGCAAT TACAGAAGAT CTTATGAGGC CTTTCTTTCC	240
CAGTGCCAAG TGGACATTCC CACCTGGGGA ACTAAACATC CTTTGGGGAT GCTTTGGCAC	300
CATAAGGTGT CAACCCTGAT TGATGAAATG GTGTCGCGTC JAATGTACCG CATCATGGAA	360
AAAGCAGGGC AGGCTGCCTG GAAACAGGTG GTGAGCGAGG CTACGCTGTC TCGCATTAGT	420
AGTTTGATG TGGTGGCTCA TTTTCAGCAT CTAGCCGCCA TTGAAGCCGA GACCTGTAAA	480

TATTTGGCCT CCCGGCTGCC CATGCTACAC AACCTGCGCA TGACAGGGTC AAATGTAACC	540
ATAGTGTATA ATAGCACTTT GAATCAGGTG TTTGCTATTT TTCCAACCCC TGGTTCCCCG	600
CCAAAGCTTC ATGATTTTCA GCAATGGTTA ATAGCTGTAC ATTCTCCAT ATTTTCCTCT	660
GTTGCAGCTT CTGTACTCT TTTTGTGTG CTGTGGTTGC GGGTTCCAAT ACTACGTACT	720
GTTTTTGGTT TCCGCTGGTT AGGGGCAATT TTTCTTTCGA ACTCACAGTG AATTACACGG	780
TGTGTCCACC TTGCCTCACC CGGCAAGCAG CCACAGAGAT CTACGAACCC GGTAGGTCTC	840
TTTGGTGCAG GATAGGGTAT GACCGATGTG GGGAGGACGA TCATGACGAG CTAGGGTTTA	900
TGATACCGCC TGGCCTCTCC AGCGAAGGCC ACTTGACTGG TGTTACGCC TGGTTGGCGT	960
TCTTGTCTTT CAGCTACAG GCCCAGTTCC ATCCCAGAT ATTCCGGATA GGGAAATGTGA	1020
GTCGAGTTTA TGTGACATC AAACATCAAC TCATCTGCGC CGAACATGAC GGGCAGAACA	1080
CCACCTTGCC TCGTCATGAC AACATTTTCA CCGTGTTCGA GACCTATTAC CAACATCAAG	1140
TCGACGGCGG CAATTGGTTT CACCTAGAAT GGCTTCGTCC CTCTTTTCC TCGTGGTTGG	1200
TTTTAAATGT CTCTTGGTTT CTCAGGCGTT CGCCTGCAA CCATGTTTCA GTTCGAGTCT	1260
TGCAGATATT AAGACCAACA CCACCGCAGC GGCAAGCTTT GCTGTCCTCC AAGACATCAG	1320
TTGCCTTAGG CATCGCGACT CGGCCTCTGA GCGGATTGCG AAAATCCCTC AGTGCCGTAC	1380
GGCGATAGGG ACACCCGTGT ATGTTACCAT CACAGCCAAT GTGACAGATG AGAATTATTT	1440
ACATTCTTCT GATCTCTCA TGCTTTCTTC TTGCCTTTTC TATGCTTCTG AGATGAGTGA	1500
AAAGGGATTT AAGGTGGTAT TTGGCAATGT GTCAGGCATC GTGGCTGTGT GTGTCAATTT	1560
TACCAGCTAC GTCCAACATG TCAAGGAGTT TACCCAACGC TCCCTGGTGG TCGACCATGT	1620
GCGGTTGCTC CATTTTATGA CACCTGAGAC CATGAGGTGG GCAACTGTTT TAGCCTGTCT	1680
TTTGTCCATT CTGTTGGCAA TTTGAATGTT TAAGTATGTT GGAGAAATGC TTGACCGCGG	1740
GCTGTTGCTC GCGATTGCTT TCTTTGTGGT GTATCGTGCC GTTCTGTTTT GCTGTGCTCG	1800
CCAACGCCAG CAACGACAGC AGCTCCCATC TACAGCTGAT TTACAACCTG ACGCTATGTG	1860
AGCTGAATGG CACAGATTGG CTAGCTAACA AATTTGATTG GGCAGTGGAG AGTTTTGTCA	1920
TCTTTCCCGT TTTGACTCAC ATTGTCTCCT ATGGTGCCCT CACTACCAGC CATTTCCCTG	1980
ACACAGTCGC TTTAGTCACT GTGTCTACCG CCGGGTTTGT TCACGGGCGG TATGTCCTAA	2040
GTAGCATCTA CGCGGTCTGT GCCCTGGCTG CGTTGACTTG CTTCTGCATT AGGTTTGCAA	2100
AGAATTGCAT GTCCTGGCGC TACGCGTGTA CCAGATATAC CAACTTTCTT CTGGACACTA	2160

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AGGGCAGACT CTATCGTTGG CGGTCGCCTG TCATCATAGA GAAAAGGGGC AAAGTTGAGG	2220
TCGAAGGTCA TCTGATCGAC CTCAAAAGAG TTGTGCTTGA TGGTTCCGTG GCAACCCCTA	2280
TAACCAGAGT TTCAGCGGAA CAATGGGGTC GTCCTTAGAT GACTTCTGTC ATGATAGCAC	2340
GGCTCCACAA AAGGTGCTTT TGGCGTTTTC TATTACCTAC ACGCCAGTGA TGATATATGC	2400
CCTAAAGGTG AGTCGCGGCC GACTGCTAGG GCTTCTGCAC CTTTGTATCT TCCTGAATTG	2460
TGCTTTCACC TTCGGGTACA TGACTTTCGC GCACTTTCAG AGTACAAATA AGGTCGCGCT	2520
CACTATGGGA GCAGTAGTTG CACTCCTTTG GGGGGTGATC TCAGCCATAG AAACCTGGAA	2580
ATTATCACC TCCAGATGCC GTTTGTGCTT GCTAGGCCGC AAGTACATTC TGGCCCTGTC	2640
CCACCACGTT GAAAGTGCCG CACGGTTTCA TCCGATTGCG GCAAATGATA ACCACGCATT	2700
TGTCGTCCGG CGTCCCGGCT CCACTACGGT CAACGGCACA TTGGTGCCCG GGTAAAAAG	2760
CCTCGTGTG GGTGGCAGAA AAGCTGTAA ACAGGGAGTG GTAAACCTTG TCAAATATGC	2820
CAAATAACAA CGGCAAGCAG ACAGAAGAGA AGAAGGGGGA TGGCCAGCCA GTCAATCAGC	2880
TGTGCCAGAT GCTGGGTAAG ATCATCGCTC AGCAAAACCA GTCCAGAGGC AAGGGACCGG	2940
GAAAGAAAAA TAAGAAGAAA AACCCGGAGA AGCCCCATTT TCCTCTAGCG ACTGAAGATG	3000
ATGTCAGACA TCACTTTACC CCTAGTGAGC GGCAATTGTG TCTGTCGTCA ATCCAGACCG	3060
CCTTTAATCA AGGCGCTGGG ACTTGCACCC TGTGAGATTC AGGGAGGATA AGTTACTG	3120
TGGAGTTTAG TTGCCTACG CATCATACTG TGCGCCTGAT CCGCGTCACA GCATCACCTT	3180
CAGCATGATG GGCTGGCATT CTTGAGGCAT CTCAGTGTG GAATTGGAAG AATGTGTGGT	3240
GAATGGCACT GATTGACATT GTGCCTCTAA GTCACCTATT CAATTAGGGC GACCGTGTGG	3300
GGGTGAGATT TAATTGGCGA GAACCATGCG GCCGAAATTA AAAAAA AAAA	3358

(2) INFORMATION FOR SEQ ID NO:2:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 768 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 1..768
- (C) IDENTIFICATION METHOD: experimental

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(D) OTHER INFORMATION: /evidence= EXPERIMENTAL
/standard_name= "VR-2332 ORF 2"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

ATG AAA TGG GGT CCA TGC AAA GCC TTT TTG ACA AAA TTG GCC AAC TTT	48
Met Lys Trp Gly Pro Cys Lys Ala Phe Leu Thr Lys Leu Ala Asn Phe	
1 5 10 15	
TTG TGG ATG CTT TCA CGG AGT TCT TGG TGT CCA TTG TTG ATA TCA TTA	96
Leu Trp Met Leu Ser Arg Ser Ser Trp Cys Pro Leu Leu Ile Ser Leu	
20 25 30	
TAT TTT TGG CCA TTT TGT TTG GCT TCA CCA TCG CCG GTT GGC TGG TGG	144
Tyr Phe Trp Pro Phe Cys Leu Ala Ser Pro Ser Pro Val Gly Trp Trp	
35 40 45	
TCT TTT GCA TCA GAT TGG TTT GCT CCG CGA TAC TCC GTA CGC GCC CTG	192
Ser Phe Ala Ser Asp Trp Phe Ala Pro Arg Tyr Ser Val Arg Ala Leu	
50 55 60	
CCA TTC ACT CTG AGC AAT TAC AGA AGA TCT TAT GAG GCC TTT CTT TCC	240
Pro Phe Thr Leu Ser Asn Tyr Arg Arg Ser Tyr Glu Ala Phe Leu Ser	
65 70 75 80	
CAG TGC CAA GTG GAC ATT CCC ACC TGG GGA ACT AAA CAT CCT TTG GGG	288
Gln Cys Gln Val Asp Ile Pro Thr Trp Gly Thr Lys His Pro Leu Gly	
85 90 95	
ATG CTT TGG CAC CAT AAG GTG TCA ACC CTG ATT GAT GAA ATG GTG TCG	336
Met Leu Trp His His Lys Val Ser Thr Leu Ile Asp Glu Met Val Ser	
100 105 110	
CGT CGA ATG TAC CGC ATC ATG GAA AAA GCA GGG CAG GCT GCC TGG AAA	384
Arg Arg Met Tyr Arg Ile Met Glu Lys Ala Gly Gln Ala Ala Trp Lys	
115 120 125	
CAG GTG GTG AGC GAG GCT ACG CTG TCT CGC ATT AGT AGT TTG GAT GTG	432
Gln Val Val Ser Glu Ala Thr Leu Ser Arg Ile Ser Ser Leu Asp Val	
130 135 140	
GTG GCT CAT TTT CAG CAT CTA GCC GCC ATT GAA GCC GAG ACC TGT AAA	480
Val Ala His Phe Gln His Leu Ala Ala Ile Glu Ala Glu Thr Cys Lys	
145 150 155 160	
TAT TTG GCC TCC CGG CTG CCC ATG CTA CAC AAC CTG CGC ATG ACA GGG	528
Tyr Leu Ala Ser Arg Leu Pro Met Leu His Asn Leu Arg Met Thr Gly	
165 170 175	
TCA AAT GTA ACC ATA GTG TAT AAT AGC ACT TTG AAT CAG GTG TTT GCT	576
Ser Asn Val Thr Ile Val Tyr Asn Ser Thr Leu Asn Gln Val Phe Ala	
180 185 190	

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ATT TTT CCA ACC CCT GGT TCC CGG CCA AAG CTT CAT GAT TTT CAG CAA	624
Ile Phe Pro Thr Pro Gly Ser Arg Pro Lys Leu His Asp Phe Gln Gln	
195 200 205	
TGG TTA ATA GCT GTA CAT TCC TCC ATA TTT TCC TCT GTT GCA GCT TCT	672
Trp Leu Ile Ala Val His Ser Ser Ile Phe Ser Ser Val Ala Ala Ser	
210 215 220	
TGT ACT CTT TTT GTT GTG CTG TGG TTG CGG GTT CCA ATA CTA CGT ACT	720
Cys Thr Leu Phe Val Val Leu Trp Leu Arg Val Pro Ile Leu Arg Thr	
225 230 235 240	
GTT TTT GGT TTC CGC TGG TTA GGG GCA ATT TTT CTT TCG AAC TCA CAG	768
Val Phe Gly Phe Arg Trp Leu Gly Ala Ile Phe Leu Ser Asn Ser Gln	
245 250 255	

(2) INFORMATION FOR SEQ ID NO:3:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 256 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

Met Lys Trp Gly Pro Cys Lys Ala Phe Leu Thr Lys Leu Ala Asn Phe	
1 5 10 15	
Leu Trp Met Leu Ser Arg Ser Ser Trp Cys Pro Leu Leu Ile Ser Leu	
20 25 30	
Tyr Phe Trp Pro Phe Cys Leu Ala Ser Pro Ser Pro Val Gly Trp Trp	
35 40 45	
Ser Phe Ala Ser Asp Trp Phe Ala Pro Arg Tyr Ser Val Arg Ala Leu	
50 55 60	
Pro Phe Thr Leu Ser Asn Tyr Arg Arg Ser Tyr Glu Ala Phe Leu Ser	
65 70 75 80	
Gln Cys Gln Val Asp Ile Pro Thr Trp Gly Thr Lys His Pro Leu Gly	
85 90 95	
Met Leu Trp His His Lys Val Ser Thr Leu Ile Asp Glu Met Val Ser	
100 105 110	
Arg Arg Met Tyr Arg Ile Met Glu Lys Ala Gly Gln Ala Ala Trp Lys	
115 120 125	
Gln Val Val Ser Glu Ala Thr Leu Ser Arg Ile Ser Ser Leu Asp Val	
130 135 140	

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Val Ala His Phe Gln His Leu Ala Ala Ile Glu Ala Glu Thr Cys Lys
 145 150 155 160

Tyr Leu Ala Ser Arg Leu Pro Met Leu His Asn Leu Arg Met Thr Gly
 165 170 175

Ser Asn Val Thr Ile Val Tyr Asn Ser Thr Leu Asn Gln Val Phe Ala
 180 185 190

Ile Phe Pro Thr Pro Gly Ser Arg Pro Lys Leu His Asp Phe Gln Gln
 195 200 205

Trp Leu Ile Ala Val His Ser Ser Ile Phe Ser Ser Val Ala Ala Ser
 210 215 220

Cys Thr Leu Phe Val Val Leu Trp Leu Arg Val Pro Ile Leu Arg Thr
 225 230 235 240

Val Phe Gly Phe Arg Trp Leu Gly Ala Ile Phe Leu Ser Asn Ser Gln
 245 250 255

(2) INFORMATION FOR SEQ ID NO:4:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 762 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 1..762
- (C) IDENTIFICATION METHOD: experimental
- (D) OTHER INFORMATION: /evidence= EXPERIMENTAL
 /standard_name= "VR-2332 ORF 3"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

ATG GTT AAT AGC TGT ACA TTC CTC CAT ATT TTC CTC TGT TGC AGC TTC	48
Met Val Asn Ser Cys Thr Phe Leu His Ile Phe Leu Cys Cys Ser Phe	
1 5 10 15	
TTG TAC TCT TTT TGT TGT GCT GTG GTT GCG GGT TCC AAT ACT ACG TAC	96
Leu Tyr Ser Phe Cys Cys Ala Val Val Ala Gly Ser Asn Thr Thr Tyr	
20 25 30	
TGT TTT TGG TTT CCG CTG GTT AGG GGC AAT TTT TCT TTC GAA CTC ACA	144
Cys Phe Trp Phe Pro Leu Val Arg Gly Asn Phe Ser Phe Glu Leu Thr	
35 40 45	

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GTG AAT TAC ACG GTG TGT CCA CCT TGC CTC ACC CGG CAA GCA GCC ACA	192
Val Asn Tyr Thr Val Cys Pro Pro Cys Leu Thr Arg Gln Ala Ala Thr	
50 55 60	
GAG ATC TAC GAA CCC GGT AGG TCT CTT TGG TGC AGG ATA GGG TAT GAC	240
Glu Ile Tyr Glu Pro Gly Arg Ser Leu Trp Cys Arg Ile Gly Tyr Asp	
65 70 75 80	
CGA TGT GGG GAG GAC GAT CAT GAC GAG CTA GGG TTT ATG ATA CCG CCT	288
Arg Cys Gly Glu Asp Asp His Asp Glu Leu Gly Phe Met Ile Pro Pro	
85 90 95	
GGC CTC TCC AGC GAA GGC CAC TTG ACT GGT GTT TAC GCC TGG TTG GCG	336
Gly Leu Ser Ser Glu Gly His Leu Thr Gly Val Tyr Ala Trp Leu Ala	
100 105 110	
TTC TTG TCC TTC AGC TAC ACG GCC CAG TTC CAT CCC GAG ATA TTC GGG	384
Phe Leu Ser Phe Ser Tyr Thr Ala Gln Phe His Pro Glu Ile Phe Gly	
115 120 125	
ATA GGG AAT GTG AGT CGA GTT TAT GTT GAC ATC AAA CAT CAA CTC ATC	432
Ile Gly Asn Val Ser Arg Val Tyr Val Asp Ile Lys His Gln Leu Ile	
130 135 140	
TGC GCC GAA CAT GAC GGG CAG AAC ACC ACC TTG CCT CGT CAT GAC AAC	480
Cys Ala Glu His Asp Gly Gln Asn Thr Thr Leu Pro Arg His Asp Asn	
145 150 155 160	
ATT TCA GCC GTG TTT CAG ACC TAT TAC CAA CAT CAA GTC GAC GGC GGC	528
Ile Ser Ala Val Phe Gln Thr Tyr Tyr Gln His Gln Val Asp Gly Gly	
165 170 175	
AAT TGG TTT CAC CTA GAA TGG CTT CGT CCC TTC TTT TCC TCG TGG TTG	576
Asn Trp Phe His Leu Glu Trp Leu Arg Pro Phe Phe Ser Ser Trp Leu	
180 185 190	
GTT TTA AAT GTC TCT TGG TTT CTC AGG CGT TCG CCT GCA AAC CAT GTT	624
Val Leu Asn Val Ser Trp Phe Leu Arg Arg Ser Pro Ala Asn His Val	
195 200 205	
TCA GTT CGA GTC TTG CAG ATA TTA AGA CCA ACA CCA CCG CAG CGG CAA	672
Ser Val Arg Val Leu Gln Ile Leu Arg Pro Thr Pro Pro Gln Arg Gln	
210 215 220	
GCT TTG CTG TCC TCC AAG ACA TCA GTT GCC TTA GGC ATC GCG ACT CGG	720
Ala Leu Leu Ser Ser Lys Thr Ser Val Ala Leu Gly Ile Ala Thr Arg	
225 230 235 240	
CCT CTG AGG CGA TTC GCA AAA TCC CTC AGT GCC GTA CGG CGA	762
Pro Leu Arg Arg Phe Ala Lys Ser Leu Ser Ala Val Arg Arg	
245 250	

(2) INFORMATION FOR SEQ ID NO:5:

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(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 254 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

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Met Val Asn Ser Cys Thr Phe Leu His Ile Phe Leu Cys Cys Ser Phe
 1           5           10           15
Leu Tyr Ser Phe Cys Cys Ala Val Val Ala Gly Ser Asn Thr Thr Tyr
          20           25           30
Cys Phe Trp Phe Pro Leu Val Arg Gly Asn Phe Ser Phe Glu Leu Thr
          35           40           45
Val Asn Tyr Thr Val Cys Pro Pro Cys Leu Thr Arg Gln Ala Ala Thr
          50           55           60
Glu Ile Tyr Glu Pro Gly Arg Ser Leu Trp Cys Arg Ile Gly Tyr Asp
65           70           75           80
Arg Cys Gly Glu Asp Asp His Asp Glu Leu Gly Phe Met Ile Pro Pro
          85           90           95
Gly Leu Ser Ser Glu Gly His Leu Thr Gly Val Tyr Ala Trp Leu Ala
          100          105          110
Phe Leu Ser Phe Ser Tyr Thr Ala Gln Phe His Pro Glu Ile Phe Gly
          115          120          125
Ile Gly Asn Val Ser Arg Val Tyr Val Asp Ile Lys His Gln Leu Ile
          130          135          140
Cys Ala Glu His Asp Gly Gln Asn Thr Thr Leu Pro Arg His Asp Asn
          145          150          155          160
Ile Ser Ala Val Phe Gln Thr Tyr Tyr Gln His Gln Val Asp Gly Gly
          165          170          175
Asn Trp Phe His Leu Glu Trp Leu Arg Pro Phe Phe Ser Ser Trp Leu
          180          185          190
Val Leu Asn Val Ser Trp Phe Leu Arg Arg Ser Pro Ala Asn His Val
          195          200          205
Ser Val Arg Val Leu Gln Ile Leu Arg Pro Thr Pro Pro Gln Arg Gln
          210          215          220
Ala Leu Leu Ser Ser Lys Thr Ser Val Ala Leu Gly Ile Ala Thr Arg
          225          230          235          240

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Pro Leu Arg Arg Phe Ala Lys Ser Leu Ser Ala Val Arg Arg
 245 250

(2) INFORMATION FOR SEQ ID NO:6:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 534 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 1..534
- (C) IDENTIFICATION METHOD: experimental
- (D) OTHER INFORMATION: /evidence= EXPERIMENTAL
 /standard_name= "VR-2332 ORF 4"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

ATG GCT TCG TCC CTT CTT TTC CTC GTG GTT GGT TTT AAA TGT CTC TTG	48
Met Ala Ser Ser Leu Leu Phe Leu Val Val Gly Phe Lys Cys Leu Leu	
1 5 10 15	
GTT TCT CAG GCG TTC GCC TGC AAA CCA TGT TTC AGT TCG AGT CTT GCA	96
Val Ser Gln Ala Phe Ala Cys Lys Pro Cys Phe Ser Ser Ser Leu Ala	
20 25 30	
GAT ATT AAG ACC AAC ACC ACC GCA GCG GCA AGC TTT GCT GTC CTC CAA	144
Asp Ile Lys Thr Asn Thr Thr Ala Ala Ala Ser Phe Ala Val Leu Gln	
35 40 45	
GAC ATC AGT TGC CTT AGG CAT CGC GAC TCG GCC TCT GAG GCG ATT CGC	192
Asp Ile Ser Cys Leu Arg His Arg Asp Ser Ala Ser Glu Ala Ile Arg	
50 55 60	
AAA ATC CCT CAG TGC CGT ACG GCG ATA GGG ACA CCC GTG TAT GTT ACC	240
Lys Ile Pro Gln Cys Arg Thr Ala Ile Gly Thr Pro Val Tyr Val Thr	
65 70 75 80	
ATC ACA GCC AAT GTG ACA GAT GAG AAT TAT TTA CAT TCT TCT GAT CTC	288
Ile Thr Ala Asn Val Thr Asp Glu Asn Tyr Leu His Ser Ser Asp Leu	
85 90 95	
CTC ATG CTT TCT TCT TGC CTT TTC TAT GCT TCT GAG ATG AGT GAA AAG	336
Leu Met Leu Ser Ser Cys Leu Phe Tyr Ala Ser Glu Met Ser Glu Lys	
100 105 110	
GGA TTT AAG GTG GTA TTT GGC AAT GTG TCA GGC ATC GTG GCT GTG TGT	384
Gly Phe Lys Val Val Phe Gly Asn Val Ser Gly Ile Val Ala Val Cys	
115 120 125	

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GTC AAT TTT ACC AGC TAC GTC CAA CAT GTC AAG GAG TTT ACC CAA CGC	432
Val Asn Phe Thr Ser Tyr Val Gln His Val Lys Glu Phe Thr Gln Arg	
130 135 140	
TCC CTG GTG GTC GAC CAT GTG CGG TTG CTC CAT TTC ATG ACA CCT GAG	480
Ser Leu Val Val Asp His Val Arg Leu Leu His Phe Met Thr Pro Glu	
145 150 155 160	
ACC ATG AGG TGG GCA ACT GTT TTA GCC TGT CTT TTT GCC ATT CTG TTG	528
Thr Met Arg Trp Ala Thr Val Leu Ala Cys Leu Phe Ala Ile Leu Leu	
165 170 175	
GCA ATT	534
Ala Ile	

(2) INFORMATION FOR SEQ ID NO:7:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 178 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

Met Ala Ser Ser Leu Leu Phe Leu Val Val Gly Phe Lys Cys Leu Leu	
1 5 10 15	
Val Ser Gln Ala Phe Ala Cys Lys Pro Cys Phe Ser Ser Ser Leu Ala	
20 25 30	
Asp Ile Lys Thr Asn Thr Thr Ala Ala Ala Ser Phe Ala Val Leu Gln	
35 40 45	
Asp Ile Ser Cys Leu Arg His Arg Asp Ser Ala Ser Glu Ala Ile Arg	
50 55 60	
Lys Ile Pro Gln Cys Arg Thr Ala Ile Gly Thr Pro Val Tyr Val Thr	
65 70 75 80	
Ile Thr Ala Asn Val Thr Asp Glu Asn Tyr Leu His Ser Ser Asp Leu	
85 90 95	
Leu Met Leu Ser Ser Cys Leu Phe Tyr Ala Ser Glu Met Ser Glu Lys	
100 105 110	
Gly Phe Lys Val Val Phe Gly Asn Val Ser Gly Ile Val Ala Val Cys	
115 120 125	
Val Asn Phe Thr Ser Tyr Val Gln His Val Lys Glu Phe Thr Gln Arg	
130 135 140	

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Ser Leu Val Val Asp His Val Arg Leu Leu His Phe Met Thr Pro Glu
 145 150 155 160

Thr Met Arg Trp Ala Thr Val Leu Ala Cys Leu Phe Ala Ile Leu Leu
 165 170 175

Ala Ile

(2) INFORMATION FOR SEQ ID NO:8:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 600 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 1..600
- (C) IDENTIFICATION METHOD: experimental
- (D) OTHER INFORMATION: /evidence= EXPERIMENTAL
 /standard_name= "VR-2332 ORF5"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

ATG TTG GAG AAA TGC TTG ACC GCG GGC TGT TGC TCG CGA TTG CTT TCT	48
Met Leu Glu Lys Cys Leu Thr Ala Gly Cys Cys Ser Arg Leu Leu Ser	
1 5 10 15	
TTG TGG TGT ATC GTG CCG TTC TGT TTT GCT GTG CTC GCC AAC GCC AGC	96
Leu Trp Cys Ile Val Pro Phe Cys Phe Ala Val Leu Ala Asn Ala Ser	
20 25 30	
AAC GAC AGC AGC TCC CAT CTA CAG CTG ATT TAC AAC TTG ACG CTA TGT	144
Asn Asp Ser Ser His Leu Gln Leu Ile Tyr Asn Leu Thr Leu Cys	
35 40 45	
GAG CTG AAT GGC ACA GAT TGG CTA GCT AAC AAA TTT GAT TGG GCA GTG	192
Glu Leu Asn Gly Thr Asp Trp Leu Ala Asn Lys Phe Asp Trp Ala Val	
50 55 60	
GAG AGT TTT GTC ATC TTT CCC GTT TTG ACT CAC ATT GTC TCC TAT GGT	240
Glu Ser Phe Val Ile Phe Pro Val Leu Thr His Ile Val Ser Tyr Gly	
65 70 75 80	
GCC CTC ACT ACC AGC CAT TTC CTT GAC ACA GTC GCT TTA GTC ACT GTG	288
Ala Leu Thr Thr Ser His Phe Leu Asp Thr Val Ala Leu Val Thr Val	
85 90 95	

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TCT ACC GCC GGG TTT GTT CAC GGG CGG TAT GTC CTA AGT AGC ATC TAC Ser Thr Ala Gly Phe Val His Gly Arg Tyr Val Leu Ser Ser Ile Tyr 100 105 110	336
GCG GTC TGT GCC CTG GCT GCG TTG ACT TGC TTC GTC ATT AGG TTT GCA Ala Val Cys Ala Leu Ala Ala Leu Thr Cys Phe Val Ile Arg Phe Ala 115 120 125	384
AAG AAT TGC ATG TCC TGG CGC TAC GCG TGT ACC AGA TAT ACC AAC TTT Lys Asn Cys Met Ser Trp Arg Tyr Ala Cys Thr Arg Tyr Thr Asn Phe 130 135 140	432
CTT CTG GAC ACT AAG GGC AGA CTC TAT CGT TGG CGG TCG CCT GTC ATC Leu Leu Asp Thr Lys Gly Arg Leu Tyr Arg Trp Arg Ser Pro Val Ile 145 150 155 160	480
ATA GAG AAA AGG GGC AAA GTT GAG GTC GAA GGT CAT CTG ATC GAC CTC Ile Glu Lys Arg Gly Lys Val Glu Val Glu Gly His Leu Ile Asp Leu 165 170 175	528
AAA AGA GTT GTG CTT GAT GGT TCC GTG GCA ACC CCT ATA ACC AGA GTT Lys Arg Val Val Leu Asp Gly Ser Val Ala Thr Pro Ile Thr Arg Val 180 185 190	576
TCA GCG GAA CAA TGG GGT CGT CCT Ser Ala Glu Gln Trp Gly Arg Pro 195 200	600

(2) INFORMATION FOR SEQ ID NO:9:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 200 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

Met Leu Glu Lys Cys Leu Thr Ala Gly Cys Cys Ser Arg Leu Leu Ser 1 5 10 15
Leu Trp Cys Ile Val Pro Phe Cys Phe Ala Val Leu Ala Asn Ala Ser 20 25 30
Asn Asp Ser Ser Ser His Leu Gln Leu Ile Tyr Asn Leu Thr Leu Cys 35 40 45
Glu Leu Asn Gly Thr Asp Trp Leu Ala Asn Lys Phe Asp Trp Ala Val 50 55 60
Glu Ser Phe Val Ile Phe Pro Val Leu Thr His Ile Val Ser Tyr Gly 65 70 75 80

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(2) INFORMATION FOR SEQ ID NO:10:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 522 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(ix) **FEATURE:**

- (A) NAME/KEY: CDS
(B) LOCATION: 1..522
(C) IDENTIFICATION METHOD: experimental
(D) OTHER INFORMATION: /evidence= EXPERIMENTAL
/standard name= "VR-2332 ORF 6"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

ATG	GGG	TCG	TCC	TTA	GAT	GAC	TTC	TGT	CAT	GAT	AGC	ACG	GCT	CCA	CAA	48
Met	Gly	Ser	Ser	Leu	Asp	Asp	Phe	Cys	His	Asp	Ser	Thr	Ala	Pro	Gln	
1				5				10					15			
AAG	GTG	CTT	TTG	GCG	TTT	TCT	ATT	ACC	TAC	ACG	CCA	GTG	ATG	ATA	TAT	96
Lys	Val	Leu	Leu	Ala	Phe	Ser	Ile	Thr	Tyr	Thr	Pro	Val	Met	Ile	Tyr	
			20					25					30			

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GCC CTA AAG GTG AGT CGC GGC CGA CTG CTA GGG CTT CTG CAC CTT TTG	144
Ala Leu Lys Val Ser Arg Gly Arg Leu Leu Gly Leu Leu His Leu Leu	
35 40 45	
ATC TTC CTG AAT TGT GCT TTC ACC TTC GGG TAC ATG ACT TTC GCG CAC	192
Ile Phe Leu Asn Cys Ala Phe Thr Phe Gly Tyr Met Thr Phe Ala His	
50 55 60	
TTT CAG AGT ACA AAT AAG GTC GCG CTC ACT ATG GGA GCA GTA GTT GCA	240
Phe Gln Ser Thr Asn Lys Val Ala Leu Thr Met Gly Ala Val Val Ala	
65 70 75 80	
CTC CTT TGG GGG GTG TAC TCA GCC ATA GAA ACC TGG AAA TTC ATC ACC	288
Leu Leu Trp Gly Val Tyr Ser Ala Ile Glu Thr Trp Lys Phe Ile Thr	
85 90 95	
TCC AGA TGC CGT TTG TGC TTG CTA GGC CGC AAG TAC ATT CTG GCC CCT	336
Ser Arg Cys Arg Leu Cys Leu Leu Gly Arg Lys Tyr Ile Leu Ala Pro	
100 105 110	
GCC CAC CAC GTT GAA AGT GCC GCA CGG TTT CAT CCG ATT GCG GCA AAT	384
Ala His His Val Glu Ser Ala Ala Arg Phe His Pro Ile Ala Ala Asn	
115 120 125	
GAT AAC CAC GCA TTT GTC GTC CGG CGT CCC GGC TCC ACT ACG GTC AAC	432
Asp Asn His Ala Phe Val Val Arg Arg Pro Gly Ser Thr Thr Val Asn	
130 135 140	
GGC ACA TTG GTG CCC GGG TTA AAA AGC CTC GTG TTG GGT GGC AGA AAA	480
Gly Thr Leu Val Pro Gly Leu Lys Ser Leu Val Leu Gly Gly Arg Lys	
145 150 155 160	
GCT GTT AAA CAG GGA GTG GTA AAC CTT GTC AAA TAT GCC AAA	522
Ala Val Lys Gln Gly Val Val Asn Leu Val Lys Tyr Ala Lys	
165 170	

(2) INFORMATION FOR SEQ ID NO:11:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 174 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

Met Gly Ser Ser Leu Asp Asp Phe Cys His Asp Ser Thr Ala Pro Gln
1 5 10 15
Lys Val Leu Leu Ala Phe Ser Ile Thr Tyr Thr Pro Val Met Ile Tyr
20 25 30

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Ala	Leu	Lys	Val	Ser	Arg	Gly	Arg	Leu	Leu	Gly	Leu	Leu	His	Leu	Leu	35	40	45
Ile	Phe	Leu	Asn	Cys	Ala	Phe	Thr	Phe	Gly	Tyr	Met	Thr	Phe	Ala	His	50	55	60
Phe	Gln	Ser	Thr	Asn	Lys	Val	Ala	Leu	Thr	Met	Gly	Ala	Val	Val	Ala	65	70	75
Leu	Leu	Trp	Gly	Val	Tyr	Ser	Ala	Ile	Glu	Thr	Trp	Lys	Phe	Ile	Thr	85	90	95
Ser	Arg	Cys	Arg	Leu	Cys	Leu	Leu	Gly	Arg	Lys	Tyr	Ile	Leu	Ala	Pro	100	105	110
Ala	His	His	Val	Glu	Ser	Ala	Ala	Arg	Phe	His	Pro	Ile	Ala	Ala	Asn	115	120	125
Asp	Asn	His	Ala	Phe	Val	Val	Arg	Arg	Pro	Gly	Ser	Thr	Thr	Val	Asn	130	135	140
Gly	Thr	Leu	Val	Pro	Gly	Leu	Lys	Ser	Leu	Val	Leu	Gly	Gly	Arg	Lys	145	150	155
Ala	Val	Lys	Gln	Gly	Val	Val	Asn	Leu	Val	Lys	Tyr	Ala	Lys			165	170	

(2) INFORMATION FOR SEQ ID NO:12:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 369 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(ix) **FEATURE:**

- (A) NAME/KEY: CDS
(B) LOCATION: 1..369
(C) IDENTIFICATION METHOD: experimental
(D) OTHER INFORMATION: /evidence= EXPERIMENTAL
/standard name= "VR-2332 ORF 7"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

ATG CCA AAT AAC AAC GGC AAG CAG ACA GAA GAG AAG AAG GGG GAT GGC 48
Met Pro Asn Asn Asn Gly Lys Gln Thr Glu Glu Lys Lys Gly Asp Gly
1 5 10 15

CAG CCA GTC AAT CAG CTG TGC CAG ATG CTG GGT AAG ATC ATC GCT CAG 96
Gln Pro Val Asn Gln Leu Cys Gln Met Leu Gly Lys Ile Ile Ala Gln
20 25 30

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CAA AAC CAG TCC AGA GGC AAG GGA CCG GGA AAG AAA AAT AAG AAG AAA	144
Gln Asn Gln Ser Arg Gly Lys Gly Pro Gly Lys Lys Asn Lys Lys Lys	
35 40 45	
AAC CCG GAG AAG CCC CAT TTT CCT CTA GCG ACT GAA GAT GAT GTC AGA	192
Asn Pro Glu Lys Pro His Phe Pro Leu Ala Thr Glu Asp Asp Val Arg	
50 55 60	
CAT CAC TTT ACC CCT AGT GAG CGG CAA TTG TGT CTG TCG TCA ATC CAG	240
His His Phe Thr Pro Ser Glu Arg Gln Leu Cys Leu Ser Ser Ile Gln	
65 70 75 80	
ACC GCC TTT AAT CAA GGC GCT GGG ACT TGC ACC CTG TCA GAT TCA GGG	288
Thr Ala Phe Asn Gln Gly Ala Gly Thr Cys Thr Leu Ser Asp Ser Gly	
85 90 95	
AGG ATA AGT TAC ACT GTG GAG TTT AGT TTG CCT ACG CAT CAT ACT GTG	336
Arg Ile Ser Tyr Thr Val Glu Phe Ser Leu Pro Thr His His Thr Val	
100 105 110	
CGC CTG ATC CGC GTC ACA GCA TCA CCC TCA GCA	369
Arg Leu Ile Arg Val Thr Ala Ser Pro Ser Ala	
115 120	

(2) INFORMATION FOR SEQ ID NO:13:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 123 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

Met Pro Asn Asn Asn Gly Lys Gln Thr Glu Glu Lys Lys Gly Asp Gly	
1 5 10 15	
Gln Pro Val Asn Gln Leu Cys Gln Met Leu Gly Lys Ile Ile Ala Gln	
20 25 30	
Gln Asn Gln Ser Arg Gly Lys Gly Pro Gly Lys Lys Asn Lys Lys Lys	
35 40 45	
Asn Pro Glu Lys Pro His Phe Pro Leu Ala Thr Glu Asp Asp Val Arg	
50 55 60	
His His Phe Thr Pro Ser Glu Arg Gln Leu Cys Leu Ser Ser Ile Gln	
65 70 75 80	
Thr Ala Phe Asn Gln Gly Ala Gly Thr Cys Thr Leu Ser Asp Ser Gly	
85 90 95	

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Arg Ile Ser Tyr Thr Val Glu Phe Ser Leu Pro Thr His His Thr Val
100 105 110

Arg Leu Ile Arg Val Thr Ala Ser Pro Ser Ala
115 120

(2) INFORMATION FOR SEQ ID NO:14:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 15101 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: Arteriviridae
- (B) STRAIN: VR-2332

(ix) FEATURE:

- (A) NAME/KEY: misc_feature
- (B) LOCATION: 7384..11775
- (C) IDENTIFICATION METHOD: experimental
- (D) OTHER INFORMATION: /evidence= EXPERIMENTAL
/label= ORF1b
/citation= ([1])

(ix) FEATURE:

- (A) NAME/KEY: misc_feature
- (B) LOCATION: 11786..12535
- (D) OTHER INFORMATION: /standard_name= "LV ORF 2"
/citation= ([1])

(ix) FEATURE:

- (A) NAME/KEY: misc_feature
- (B) LOCATION: 212..7402
- (D) OTHER INFORMATION: /standard_name= "LV ORF 1a"
/citation= ([1])

(ix) FEATURE:

- (A) NAME/KEY: misc_feature
- (B) LOCATION: 12394..13191
- (D) OTHER INFORMATION: /standard_name= "LV ORF 3"
/citation= ([1])

(ix) FEATURE:

- (A) NAME/KEY: misc_feature
- (B) LOCATION: 12936..13487
- (D) OTHER INFORMATION: /standard_name= "LV ORF 4"
/citation= ([1])

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(ix) FEATURE:

- (A) NAME/KEY: misc_feature
- (B) LOCATION: 13484..14089
- (D) OTHER INFORMATION: /standard_name= "LV ORF 5"
/citation= ([1])

(ix) FEATURE:

- (A) NAME/KEY: misc_feature
- (B) LOCATION: 14077..14598
- (D) OTHER INFORMATION: /standard_name= "LV ORF 6"
/citation= ([1])

(ix) FEATURE:

- (A) NAME/KEY: misc_feature
- (B) LOCATION: 14588..14974
- (D) OTHER INFORMATION: /standard_name= "LV ORF 7"
/citation= ([1])

(x) PUBLICATION INFORMATION:

- (A) AUTHORS: Meulenberg, J. J.M.
Hulst, M. M.
de Veijer, E. J.
Moonen, P. L.
den Besten, A.
de Kluyver, E. P.
Wensvoort, G.
Moormann, R. J.
- (B) TITLE: Lelystad virus, the causative agent of
procine epidemic abortion and respiratory
syndrome (PEARS) is related to LDV and EAV.
- (C) JOURNAL: Virology
- (D) VOLUME: 192
- (F) PAGES: 62-72
- (G) DATE: 1993
- (K) RELEVANT RESIDUES IN SEQ ID NO:14: FROM 1 TO 15101

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

GGGTATTCCC CCTACATACA CGACACTTCT AGTGTGTTGTG TACCTTGGAG GCGTGGGTAC	60
AGCCCCGCCC CACCCCTTGG CCCCTGTTCT AGCCCAACAG GTATCCTTCT CTCTCGGGGC	120
GAGTGCGCCG CCTGTGCTC CTTGCAGCG GGAAGGACCT CCCGAGTATT TCCGGAGAGC	180
ACCTGCTTTA CGGGATCTCC ACCCTTTAAC CATGTCTGGG ACGTTCTCCC GGTGCATGTG	240
CACCCCGGCT GCCCGGGTAT TTTGGAACGC CGGCCAAGTC TTTGCACAC GGTGTCTCAG	300
TGCGCGGTCT CTTCTCTCTC CAGAGCTTCA GGACACTGAC CTCGGTGCAG TTGGCTTGTT	360
TTACAAGCCT AGGGACAAGC TTTACTGGAA AGTCCCTATC GGCATCCCTC AGGTGGAATG	420
TACTCCATCC GGGTGCTGTT GGCTCTCAGC TGTTCCTTCCCT TTGGCGCGTA TGACCTCCGG	480

CAATCACAAC TTCCTCCAAC GACTTGTGAA GGTTGCTGAT GTTTTGTACC GTGACGGTTG 540
CTTGGCACCT CGACACCTTC GTGAACTCCA AGTTTACGAG CGCGGCTGCA ACTGGTACCC 600
GATCACGGGG CCCGTGCCCC GGATGGGTTT GTTTGCGAAC TCCATGCACG TATCCGACCA 660
GCCGTTCCCT GGTGCCACCC ATGTGTTGAC TAACTCGCCT TTGCCTCAAC AGGCTTGTCTG 720
GCAGCCGTTT TGTCCATTG AGGAGGCTCA TTCTAGCGTG TACAGGTGGA AGAAATTTGT 780
GGTTTTCACG GACTCCTCCC TCAACGGTCG ATCTCGCATG ATGTGGACGC CGGAATCCGA 840
TGATTACGCC GCCCTGGAGG TACTACCGCC TGAGTTAGAA CGTCAGGTCTG AAATCCTCAT 900
TCGGAGTTTT CCTGCTCATC ACCCTGTCGA CCTGGCCGAC TGGGAGCTCA CTGAGTCCCC 960
TGAGAACGGT TTTTCCTTCA ACACGTCTCA TTCTTGCGGT CACCTTGTCC AGAACCCCGA 1020
CGTGTTTGAT GGCAAGTGCT GGCTCTCCTG CTTTGTGGC CAGTCGGTCG AAGTGCCTG 1080
CCATGAGGAA CATCTAGCTG ACGCCTTCGG TTACCAAACC AAGTGGGGCG TGCATGGTAA 1140
GTACCTCCAG CGCAGGCTTC AAGTTCGCGG CATTCTGTCT STAGTCGATC CTGATGGTCC 1200
CATTACGTT GAAGCGCTGT CTTGCCCCCA GTCTTGATC AGGCACCTGA CTCTGGATGA 1260
TGATGTCACC CCAGGATTCTG TTCGCCTGAC ATCCCTTCGC ATTGTGCCGA ACACAGAGCC 1320
TACCACTTCC CGGATCTTTC GGTTCGGAGC GCATAAGTGG TATGGCGCTG CCGGCAAACG 1380
GGCTCGTGCT AAGCGTGCCG CTAAAAGTGA GAAGGATTCTG GCTCCCACCC CCAAGGTTGC 1440
CCTGCCGGTC CCCACCTGTG GAATTACCAC CTACTCTCCA CCGACAGACG GGTCTTGTGG 1500
TTGGCATGTC CTTGCCGCCA TAATGAACCG GATGATAAAT GGTGACTTCA CGTCCCCTCT 1560
GACTCAGTAC AACAGACCAG AGGATGATTG GGCTTCTGAT TATGATCTTG TTCAGGCGAT 1620
TCAATGTCTA CGACTGCCTG CTACCGTGGT TCGGAATCGC GCCTGTCCTA ACGCCAAGTA 1680
CCTTATAAAA CTTAACGGAG TTTACTGGGA GGTAGAGGTG AGGTCTGGAA TGGCTCCTCG 1740
CTCCCTTTCT CGTGAATGTG TGGTTGGCGT TTGCTCTGAA GGCTGTGTCTG CACCGCCTTA 1800
TCCAGCAGAC GGGCTACCTA AACGTGCACT CGAGGCCTTG GCGTCTGCTT ACAGACTACC 1860
CTCCGATTGT GTTAGCTCTG GTATTGCTGA CTTTCTTGCT AATCCACCTC CTCAGGAATT 1920
CTGGACCCTC GACAAAATGT TGACCTCCCC GTCACCAGAG CGGTCCGGCT TCTCTAGTTT 1980
GTATAAATTA CTATTAGAGG TTGTTCCGCA AAAATGCGGT GCCACGGAAG GGGCTTTTCA 2040
CTATGCTGTT GAGAGGATGT TGAAGGATTG TCCGAGCTCC AACAGGCCA TGGCCCTTCT 2100
GGCAAAAATT AAAGTTCCAT CCTCAAAGGC CCCGTCTGTG TCCCTGGACG AGTGTTCCTC 2160

TACGGATGTT TTAGCCGACT TCGAGCCAGC ATCTCAGGAA AGGCCCCAAA GTTCCGGCGC	2220
TGCTGTTGTC CTGTGTTTAC CGGATGCAAA AGAGTTCGAG GAAGCAGCCC CGGAAGAAGT	2280
TCAAGAGAGT GGCCACAAGG CCGTCCACTC TGCACTCCTT GCCGAGGGTC CTAACAATGA	2340
GCAGGTACAG GTGGTTGCCG GTGAGCAACT GAAGCTCGGC GGTGTGGTT TGGCAGTCGG	2400
GAATGCTCAT GAAGGTGCTC TGGTCTCAGC TGGTCTAATT AACCTGGTAG GCGGGAATTT	2460
GTCCCCCTCA GACCCCATGA AAGAAAACAT GCTCAATAGC CGGGAAGACG AACCCTGGA	2520
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CCCAGGTTCT GATGCCGGTG CCTCCCCGT CACCGTTCGA GAATTTGTCC CGACGGGGCC	2640
TATACTCTGT CATGTTGAGC ACTGCGGCAC GGAGTCGGGC GACAGCAGTT CGCCTTTGGA	2700
TCTATCTGAT GCGCAAACCC TGGACCAGCC TTTAAATCTA TCCCTGGCCG CTTGGCCAGT	2760
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CAGCTCTGTC ATCGAGTTTG ACCGGACAAA AGATGCTCCG GTGGTTGACG CCCCTGTCTG	2940
CTTGACGACT TCGAACGAGG CCCTCTCTGT AGTCGATCCT TTCGAATTTG CCGAACTCAA	3000
GCGCCCGCGT TTCTCCGCAC AAGCCTTAAT TGACCGAGGC GGTCCACTTG CCGATGTCCA	3060
TGCAAAAATA AAGAACCGGG TATATGAACA GTGCCTCCAA GCTTGTGAGC CCGGTAGTCG	3120
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TGACATGATT CAAGACACAC CGCCTCCTGT TCCCAGGAAG AACCGAGCTA GTGACAATGC	3300
CGGCCTGAAG CAACTGGTGG CACAGTGGA TAGGAAATTG AGTGTGACCC CCCCCCAA	3360
ACCGGTTGGG CCAGTGCTTG ACCAGATCGT CCCTCCGCCT ACGGATATCC AGCAAGAAGA	3420
TGTCACCCCC TCCGATGGGC CACCCCATGC GCCGGATTTT CCTAGTCGAG TGAGCACGGG	3480
CGGGAGTTGG AAAGGCCTTA TGCTTTCCGG CACCCGTCTC GCGGGGTCTA TCAGCCAGCG	3540
CCTTATGACA TGGGTTTTTG AAGTTTCTC CCACCTCCCA GCTTTTATGC TCACACTTTT	3600
CTCGCCGCGG GGCTCTATGG CTCCAGGTGA TTGGTTGTTT GCAGGTGTCG TTTTACTTGC	3660
TCTCTTGCTC TGTCGTTCTT ACCCGATACT CGGATGCCTT CCCTTATTGG GTGTCTTTTC	3720
TGGTCTTTG CGGCGTGTTC GTCTGGGTGT TTTTGGTTCT TGGATGGCTT TTGCTGTATT	3780
TTTATTCTCG ACTCCATCCA ACCCAGTCGG TTCTTCTTGT GACCACGATT CGCCGGAGTG	3840

TCATGCTGAG CTTTTGGCTC TTGAGCAGCG CCAACTTTGG GAACCTGTGC GCGGCCTTGT	3900
GGTCGGCCCC TCAGGCCTCT TATGTGTCAT TCTTGGCAAG TTACTCGGTG GGTCACGTTA	3960
TCTCTGGCAT GTTCTCCTAC GTTTATGCAT GCTTGCAGAT TTGGCCCTTT CTCTTGTTTA	4020
TGTGCTGTCC CAGGGGCGTT GTCACAAGTG TTGGGGAAAG TGTATAAGGA CAGCTCCTGC	4080
GGAGGTGGCT CTTAATGTAT TTCCTTTCTC GCGCGCCACC CGTGTCTCTC TTGTATCCTT	4140
GTGTGATCGA TTCCAAACGC CAAAAGGGGT TGATCCTGTG CACTTGGCAA CGGGTTGGCG	4200
CGGGTGCTGG CGTGGTGAGA GCCCCATCCA TCAACCACAC CAAAAGCCCA TAGCTTATGC	4260
CAATTTGGAT GAAAAGAAAA TGTCTGCCCA AACGGTGGTT GCTGTCCCAT ACGATCCCAG	4320
TCAGGCTATC AAATGCCTGA AAGTTCTGCA GGCGGGAGGG GCCATCGTGG ACCAGCCTAC	4380
ACCTGAGGTC GTTCGTGTGT CCGAGATCCC CTCTCAGCC CCATTTTCC CAAAAGTTCC	4440
AGTCAACCCA GATTGCAGGG TTGTGGTAGA TTCGGACACT TTTGTGGCTG CGGTTGCTG	4500
CGGTTACTCG ACAGCACAAC TGGTTCTGGG CCGGGGCAAC TTTGCCAAGT TAAATCAGAC	4560
CCCCCCCAGG AACTCTATCT CCACCAAAC GACTGGTGGG GCCTCTTACA CCCTTGCTGT	4620
GGCTCAAGTG TCTGCGTGA CTCTTGTCA TTTCATCCTC GGTCTTGGT TCACATCACC	4680
TCAAGTGTGT GGCCGAGGAA CCGCTGACCC ATGGTGTTC AATCCTTTT CATATCCTAC	4740
CTATGGCCCC GGAGTTGTGT GCTCCTCTCG ACTTGTGTG TCTGCCGACG GGGTCACCCT	4800
GCCATTGTTT TCAGCCGTGG CACAACCTC CGGTAGAGAG GTGGGGATT TTATTTTGGT	4860
GCTCGTCTCC TTGACTGCTT TGGCCACCG CATGGCTCTT AAGGCAGACA TGTTAGTGGT	4920
CTTTTCGGCT TTTTGTGCTT ACGCCTGGCC CATGAGCTCC TGGTTAATCT GCTTCTTTCC	4980
TATACTCTTG AAGTGGGTTA CCCTTCACCC TCTTACTATG CTTTGGGTGC ACTCATTCTT	5040
GGTGTTTTGT CTGCCAGCAG CCGGCATCCT CTCACTAGGG ATAAGTGGCC TTCTTTGGGC	5100
AATTGGCCGC TTTACCCAGG TTGCCGGAAT TATTACACCT TATGACATCC ACCAGTACAC	5160
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CGTCCGAGA GCTGCTTTAA CTGGGCGAAC TTTAATCTT ACCCCGTCTG CAGTTGGATC	5280
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CTCTTCCCTT GGTTCGGAG GGGTTTTTAC CATTGATGGC AGAAGAACTG TCGTCACTGC	5400
TGCCCATGTG TTGAACGGCG ACACAGCTAG AGTCACCGGC GACTCCTACA ACCGCATGCA	5460
CACTTCAAG ACCAATGGTG ATTATGCCTG GTCCCATGCT GATGACTGGC AGGGCGTTGC	5520

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GCTCTCTGAC CTTTCCAGAC ATTTTGCAGG CCCAAGCGTT CCTCTTGGGG ACATTAAATT	5820
GAGTCCGGCC ATCATCCCTG ATGTAACATC CATTCCGAGT GACTTGGCAT CGCTCCTAGC	5880
CTCCGTCCCT GTAGTGAAG GCGGCCTCTC GACCGTTCAA CTTTGTGTG TCTTTTTCCT	5940
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GAATGAAATT CTTCCAGCAG TTTTGGTCCG AGCCGTGTTT TCTTTTGCAC TCTTTGTGCT	6060
TGCATGGGCC ACCCCCTGGT CTGCACAGGT GTTGATGATT AGACTCCTCA CGGCATCTCT	6120
CAACCGCAAC AAGCTTTCTC TGGCGTTCTA CGCACTCGGG GGTGTCGTCG GTTTGGCAGC	6180
TGAAATCGGG ACTTTTGCTG GCAGATTGTC TGAATTGTCT CAAGCTCTTT CGACATACTG	6240
CTTCTTACCT AGGGTCCTTG CTATGACCAG TTGTGTTCCC ACCATCATCA TTGGTGGACT	6300
CCATACCCTC GGTGTGATTG TGTGGTTATT CAAATACCGG TGCCTCCACA ACATGCTGGT	6360
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CCTGCGCCAA GAGTTGGCCT CTCTAGTTCA GATTGACAAA ATGAAAGGAG TTTTGTCCAA	6660
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GCTTGGGCAA CATCCTCAGG GATCCATCCT CGATATTAAT GTGGGGACTG AAAGGAAAAC	6780
TGTGTCCGTG CAAGAGACCC GGAGCCTAGG CGGCTCCAAA TTCAGTGTTT GTACTGTCGT	6840
GTCCAACACA CCCGTGGACG CCTTGACCGG CATCCCACTC CAGACACCAA CCCCTCTTTT	6900
TGAGAATGGT CCGCGTCATC GCAGCGAGGA AGACGATCTT AAAGTCGAGA GGATGAAGAA	6960
AACTGTGTA TCCCTCGGCT TCCACAACAT CAATGGCAAA GTTACTGCA AAATTGGGA	7020
CAAGTCTACC GGTGACACCT TTACACGGA TGATTCCCGG TACACCCAAG ACCATGCTTT	7080
TCAGGACAGG TCAGCCGACT ACAGAGACAG GGACTATGAG GGTGTGCAAA CCACCCCCCA	7140
ACAGGGATTT GATCCAAAGT CTGAAACCCC TGTTGGCACT GTTGTGATCG GCGGTATTAC	7200

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TGAAGCTGCC AAGCTGTCCC TTGAGCAAGC TCTCGCTGGG ATGGGCCAAA CTTGCGACCT	7320
TACAGCTGCC GAGGTGGAAA AGCTAAAGCG CATCATTAGT CAACTCCAAG GTTTGACCAC	7380
TGAACAGGCT TTAAACTGTT AGCCGCCAGC GGCTTGACCC GCTGTGGCCG CGGCGGCCTA	7440
GTGTGTGACTG AAACGGCGGT AAAAATTATA AAATACCACA GCAGAACTTT CACCTTAGGC	7500
CCTTTAGACC TAAAAGTCAC TTCCGAGGTG GAGGTAAAGA AATCAACTGA GCAGGGCCAC	7560
GCTGTGTGG CAAACTTATG TTCCGGTGTC ATCTTGATGA GACCTCACCC ACCGTCCCTT	7620
GTCGACGTTT TTCTGAAACC CGGACTTGAC ACAATACCCG GCATTCAACC AGGGCATGGG	7680
GCCGGGAATA TGGGCGTGA CGGTCTATT TGGGATTTG AAACCGCACC CACAAAGGCA	7740
GAACTCGAGT TATCCAAGCA AATAATCCAA GCATGTGAAG TTAGGCGCGG GGACGCCCCG	7800
AACCTCCAAC TCCCTTACAA GCTCTATCCT GTTAGGGGGG ATCCTGAGCG GCATAAAGGC	7860
CGCCTTATCA ATACCAGGTT TGGAGATTTA CTTACAAAA CTCCTCAAGA CACCAAGTCC	7920
GCAATCCACG CGGCTTGTG CCTGCACCCC AACGGGGCCC CCGTGTCTGA TGGTAAATCC	7980
ACACTAGGTA CCACTCTTCA ACATGGTTTC GAGCTTTATG TCCCTACTGT GCCCTATAGT	8040
GTCATGGAGT ACCTTGATTC ACGCCCTGAC ACCCCTTTTA TGTGTACTAA ACATGGCACT	8100
TCCAAGGCTG CTGCAGAGGA CCTCCAAAA TACGACCTAT CCACCCAAGG ATTTGTCCTG	8160
CCTGGGGTCC TACGCCTAGT ACGCAGATTC ATCTTTGGCC ATATTGGTAA GGCGCCGCCA	8220
TTGTTCTCC CATCAACCTA TCCCGCCAAG AACTCTATGG CAGGGATCAA TGGCCAGAGG	8280
TTCCCAACAA AGGACGTTCA GAGCATACCT GAAATTGATG AAATGTGTGC CCGCGCTGTC	8340
AAGGAGAATT GGCAAACTGT GACACCTTGC ACCCTCAAGA AACAGTACTG TTCCAAGCCC	8400
AAAACCAGGA CCATCCTGGG CACCAACAAC TTTATTGCCT TGGCTCACAG ATCGGCGCTC	8460
AGTGGTGTC CCCAGGCATT CATGAAGAAG GCTTGAAGT CCCCAATTGC CTTGGGGAAA	8520
AACAAATTCA AGGAGCTGCA TTGCACTGTC GCCGGCAGGT GTCTTGAGGC CGACTTGGCC	8580
TCCTGTGACC GCAGCACCCC CGCCATTGTA AGATGGTTTG TTGCCAACCT CCTGTATGAA	8640
CTTGCAAGAT GTGAAGAGTA CTTGCCTAGC TATGTGCTTA ATTGCTGCCA TGACCTCGTG	8700
GCAACACAGG ATGGTGCCTT CACAAAACGC GGTGGCCTGT CGTCCGGGGA CCCCGTCACC	8760
AGTGTGTCCA ACACCGTATA TTCCTGGTA ATTTATGCCC AGCACATGGT ATTGTCGGCC	8820
TTGAAAATGG GTCATGAAAT TGGTCTTAAG TTCCTCGAGG AACAGCTCAA GTTCGAGGAC	8880

CTCCTTGAAA TTCAGCCTAT GTTGGTATAC TCTGATGATC TTGTCTTGTA CGCTGAAAGA	8940
CCCACATTTT CCAATTACCA CTGGTGGGTC GAGCACCTTG ACCTGATGCT GGGTTTCAGA	9000
ACGGACCCAA AGAAAACCGT CATAACTGAT AAACCCAGCT TCCTCGGCTG CAGAATTGAG	9060
GCAGGGCGAC AGCTAGTCCC CAATCGCGAC CGCATCCTGG CTGCTCTTGC ATATCACATG	9120
AAGGCGCAGA ACGCCTCAGA GTATTATGCG TCTGCTGCCG CAATCCTGAT GGATTTCATG	9180
GCTTGCAATTG ACCATGACCC TGAGTGGTAT GAGGACCTCA TCTGCGGTAT TGCCCGGTGC	9240
GCCCGCCAGG ATGGTTATAG CTTCCCAGGT CCGGCATTTT TCATGTCCAT GTGGGAGAAG	9300
CTGAGAAGTC ATAATGAAGG GAAGAAATTC CGCCACTGCG GCATCTGCGA CGCCAAAGCC	9360
GACTATGCGT CCGCCTGTGG GCTTGATTG TGTTTGTTC ATTGCGACTT TCATCAACAC	9420
TGCCCTGTCA CTCTGAGCTG CGGTCACCAT GCCGGTTCAG AGGAATGTTC GCAGTGTCAG	9480
TCACCTGTTG GGGCTGGCAG ATCCCCTCTT GATGCCGTGC TAAAACAAAT TCCATACAAA	9540
CCTCCTCGTA CTGTCATCAT GAAGGTGGGT AATAAAACAA CGGCCCTCGA TCCGGGGAGG	9600
TACCAGTCCC GTCGAGGTCT CGTTGCAGTC AAGAGGGGTA TTGCAGGCAA TGAAGTTGAT	9660
CTTTCTGATG GGGACTACCA AGTGGTGCCT CTTTGTCCGA CTTGCAAAGA CATAAACATG	9720
GTGAAGGTGG CTTGCAATGT ACTACTCAGC AAGTTCATAG TAGGGCCACC AGGTTCCGGA	9780
AAGACCACCT GGCTACTGAG TCAAGTCCAG GACGATGATG TCATTACAC ACCCACCCT	9840
CAGACTATGT TTGATATAGT CAGTGCTCTC AAAGTTTGCA GGTATTCCAT TCCAGGAGCC	9900
TCAGGACTCC CTTTCCCACC ACCTGCCAGG TCCGGGCCGT GGGTTAGGCT TATTGCCAGC	9960
GGGCACGTCC CTGGCCGAGT ATCATACCTC GATGAGGCTG GATATTGTAA TCATCTGGAC	10020
ATTCTTAGAC TGCTTTCCAA AACACCCCTT GTGTGTTTG GTGACCTTCA GCAACTTCAC	10080
CCTGTCGGCT TTGATTCTA CTGTTATGTG TTCGATCAGA TGCCTCAGAA GCAGCTGACC	10140
ACTATTTACA GATTTGGCCC TAACATCTGC GCACGCATCC AGCCTTGTTA CAGGGAGAAA	10200
CTTGAATCTA AGGCTAGGAA CACTAGGGTG GTTTTACCA CCCGGCCTGT GGCCTTTGGT	10260
CAGGTGCTGA CACCATACCA TAAAGATCGC ATCGGCTCTG CGATAACCAT AGATTTCATC	10320
CAGGGGGCCA CCTTTGATAT TGTGACATTG CATCTACCAT CGCCAAAGTC CCTAAATAAA	10380
TCCCGAGCAC TTGTAGCCAT CACTCGGGCA AGACACGGGT TGTTCATTTA TGACCCTCAT	10440
AACCAGCTCC AGGAGTTTTT CAACTTAACC CTTGAGCGCA CTGATTGTAA CCTTGTGTTT	10500
AGCCGTGGGG ATGAGCTGGT AGTTCTGAAT GCGGATAATG CAGTCACAAC TGTAGCGAAG	10560

GCCCTTGAGA CAGGTCCATC TCGATTTCTGA GTATCAGACC CGAGGTGCAA GTCTCTCTTA 10620
GCCGCTTGTT CGGCCAGTCT GGAAGGGAGC TGTATGCCAC TACCGCAAGT GGCACATAAC 10680
CTGGGGTTTT ACTTTTCCCC GGACAGTCCA ACATTGACAC CTCTGCCAAA AGAGTTGGCG 10740
CCACATTGGC CAGTGGTTAC CCACCAGAAT AATCGGGCGT GGCCTGATCG ACTTGTGCGT 10800
AGTATGCGCC CAATTGATGC CCGCTACAGC AAGCCAATGG TCGGTGCAGG GTATGTGGTC 10860
GGGCCGTCCA CCTTTCTTGG TACTCCTGGT GTGGTGTCTAT ACTATCTCAC ACTATACATC 10920
AGGGGTGAGC CCCAGGCCTT GCCAGAAACA CTCGTTTCAA CAGGGCGTAT AGCCACAGAT 10980
TGTCGGGAGT ATCTCGACGC GGCTGAGGAA GAGGCAGCAA AAGAACTCCC CCACGCATTC 11040
ATTGGCGATG TCAAAGGTAC CACGTTTGGG GGGTGTCTATC ACATTACATC AAAATACCTA 11100
CCTAGGTCCC TGCCTAAGGA CTCTGTTGCC GTAGTTGGAG TAAGTTCGCC CGGCAGGGCT 11160
GCTAAAGCCG TGTGCACTCT CACCGATGTG TACCTCCCCG AACTCCGGCC ATATCTGCAA 11220
CCTGAGACGG CATCAAAATG CTGGAACTC AAATTAGACT TCAGGGACGT CCGACTAATG 11280
GTCTGGAAG GAGCCACCGC CTATTTCCAG TTGGAAGGGC TTACATGGTC GGCCTGCCC 11340
GACTATGCCA GGTTTATTCA GCTGCCCAAG GATGCCGTTG TATACATTGA TCCGTGTATA 11400
GGACCGGCAA CAGCCAACCG TAAGGTCGTG CGAACCACAG ACTGGCGGGC CGACCTGGCA 11460
GTGACACCGT ATGATTACGG TGCCAGAAC ATTTTGACAA CAGCCTGGTT CGAGGACCTC 11520
GGGCCGCAGT GGAAGATTTT GGGGTTGCAG CCCTTTAGGC GAGCATTGTTG CTTTGAAAAC 11580
ACTGAGGATT GGGCAATCCT TGACGCGCTG ATGAATGACG GCAAGGACTA CACTGACTAT 11640
AACTGGAAC GTGTTTCGAGA ACGCCACAC GCCATCTACG GGCCTGCTCG TGACCATACG 11700
TATCATTTTG CCCCTGGCAC AGAATTGCAG GTAGAGCTAG GTAAACCCCG GCTGCCGCCT 11760
GGGCAAGTGC CGTGAATTCG GGGTGATGCA ATGGGGTCAC TGTGGAGTAA AATCAGCCAG 11820
CTGTTCTGTTG ACGCCTTCAC TGAGTTCCTT GTTAGTGTGG TTGATATTGC CATTTTCCTT 11880
GCCATACTGT TTGGGTTTAC CGTCGCAGGA TGGTACTGTTG TCTTCTTCT CAGAGTGGTT 11940
TGCTCCGCGC TTCTCCGTTT GCGCTCTGCC ATTCACTCTC CCGAACTATC GAAGGTCCTA 12000
TGAAGGCTTG TTGCCCAACT GCAGACCGGA TGTCCACAA TTTGCAGTCA AGCACCATT 12060
GGGTATGTTT TGGCACATGC GAGTTTCCCA CTTGATTGAT GAGATGGTCT CTCGTGCGAT 12120
TTACCAGACC ATGGAACATT CAGGTCAAGC GGCCTGGAAG CAGGTGGTTG GTGAGGCCAC 12180
TCTCAGGAAG CTGTCAGGGC TCGATATAGT TACTCATTTC CAACACCTGG CCGCAGTGGA 12240

GGCGGATTCT TGCCGCTTTC TCAGCTCAGC ACTCGTGATG CTAAAAAATC TTGCCGTTGG	12300
CAATGTGAGC CTACAGTACA ACACCACGTT GGACCGCGTT GAGCTCATCT TCCCCACGCC	12360
AGGTACGAGG CCCAAGTTGA CCGATTTCAG ACAATGGCTC ATCAGTGTGC ACGCTTCCAT	12420
TTTTTCCTCT GTGGCTTCAT CTGTTACCTT GTTCATAGTG CTTTGGCTTC GAATTCCAGC	12480
TCTACGCTAT GTTTTTGTT TCCATTGGCC CACGGCAACA CATCATTCGA GCTGACCATC	12540
AACTACACCA TATGCATGCC CTGTTCTACC AGTCAAGCGG CTCGCCAAAG GCTCGAGCCC	12600
GGTCGTAACA TGTGGTGCAA AATAGGGCAT GACAGGTGTG AGGAGCGTGA CCATGATGAG	12660
TTGTTAATGT CCATCCCGTC CGGTACGAC AACCTCAAAC TTGAGGGTTA TTATGCTTGG	12720
CTGGCTTTTT TGTCCTTTTC CTACGCGGCC CAATTCCATC CGGAGTTGTT CGGGATAGGG	12780
AATGTGTCGC GCGTCTTCGT GGACAAGCGA CACCAGTTCA TTTGTGCCGA GCATGATGGA	12840
CACAATTCAA CCGTATCTAC CGGACACAAC ATCTCCGCAT TATATGCGGC ATATTACCAC	12900
CACCAAATAG ACGGGGGCAA TTGGTTCCAT TTGGAATGGC TGCGGCCACT CTTTTCTTCC	12960
TGGCTGGTGC TCAACATATC ATGTTTTCTG AGGCGTTCGC CTGTAAGCCC TGTTTCTCGA	13020
CGCATCTATC AGATATTGAG ACCAACACGA CCGCGGCTGC CGGTTTCATG GTCCTTCAGG	13080
ACATCAATTG TTTCCGACCT CACGGGGTCT CAGCAGCGCA AGAGAAAATT TCCTTCGGAA	13140
AGTCGTCCCA ATGTCGTGAA GCCGTCGGTA CTCCCCAGTA CATCACGATA ACGGCTAACG	13200
TGACCGACGA ATCATACTTG TACAACGCGG ACCTGCTGAT GCTTTCTGCG TGCCTTTTCT	13260
ACGCCTCAGA AATGAGCGAG AAAGGCTTCA AAGTCATCTT TGGGAATGTC TCTGGCGTTG	13320
TTTCTGCTTG TGTCAAATTC ACAGATTATG TGGCCCATGT GACCCAACAT ACCCAGCAGC	13380
ATCATCTGGT AATTGATCAC ATTCGGTTGC TGCATTTCTT GACACCATCT GCAATGAGGT	13440
GGGCTACAAC CATTGCTTGT TTGTTGCGCA TTCTCTTGGC AATATGAGAT GTTCTCACAA	13500
ATTGGGGCGT TTCTTGACTC CGCACTCTTG CTCTGCTGGG CTTTTTTTGC TGTGTACCGG	13560
CTTGCTCTGG TCCTTTGCCG ATGGCAACGG CGACAGCTCG ACATACCAAT ACATATATAA	13620
CTTGACGATA TGCAGAGTGA ATGGGACCGA CTGGTTGTCC AGCCATTTTG GTTGGGCAGT	13680
CGAGACCTTT GTGCTTTACC CGGTTGCCAC TCATATCCTC TCACTGGGTT TTCTCACAA	13740
AAGCCATTTT TTTGACGCGC TCGGTCTCGG CGCTGTATCC ACTGCAGGAT TTGTTGGCGG	13800
GCGGTACGTA CTCTGCAGCG TCTACGGCGC TTGTGCTTTC GCAGCGTTCG TATGTTTTGT	13860
CATCCGTGCT GCTAAAAATT GCATGGCCTG CCGCTATGCC CGTACCCGGT TTACCAACTT	13920

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CATTGTGGAC GACCGGGGGA GAGTTCATCG ATGGAAGTCT CCAATAGTGG TAGAAAAATT	13980
GGGCAAAGCC GAAGTCGATG GCAACCTCGT CACCATCAAA CATGTCGTCC TCGAAGGGGT	14040
TAAAGCTCAA CCCTTGACGA GGACTTCGGC TGAGCAATGG GAGGCCTAGA CGATTTTTGC	14100
AACGATCCTA TCGCCGCACA AAAGCTCGTG CTAGCCTTTA GCATCACATA CACACCTATA	14160
ATGATATACG CCCTTAAGGT GTCACGCGGC CGACTCCTGG GGCTGTTGCA CATCCTAATA	14220
TTTCTGAACT GTTCCTTTAC ATTCGATAC ATGACATATG TGCATTTTCA ATCCACCAAC	14280
CGTGTGCGAC TTACCCTGGG GGCTGTTGTC GCCCTTCTGT GGGGTGTTTA CAGCTTCACA	14340
GAGTCATGGA AGTTTATCAC TTCCAGATGC AGATTGTGTT GCCTTGCCCG GCGATACATT	14400
CTGGCCCCTG CCCATCACGT AGAAAGTGCT GCAGGTCTCC ATTCAATCTC AGCGTCTGGT	14460
AACCGAGCAT ACGCTGTGAG AAAGCCCGGA CTAACATCAG TGAACGGCAC TCTAGTACCA	14520
GGACTTCGGA GCCTCGTGCT GGGCGGCAAA CGAGCTGTTA AACGAGGAGT GGTTAACCTC	14580
GTCAAGTATG GCCGGTAAAA ACCAGAGCCA GAAGAAAAAG AAAAGTACAG CTCCGATGGG	14640
GAATGGCCAG CCAGTCAATC AACTGTGCCA GTTGCTGGGT GCAATGATAA AGTCCCAGCG	14700
CCAGCAACCT AGGGGAGGAC AGGCCAAAAA GAAAAAGCCT GAGAAGCCAC ATTTTCCCCT	14760
GGCTGCTGAA GATGACATCC GGCACCACCT CACCCAGACT GAACGCTCCC TCTGCTTGCA	14820
ATCGATCCAG ACGGCTTTCA ATCAAGGCGC AGGAACTGCG TCGCTTTCAT CCAGCGGGAA	14880
GGTCAGTTTT CAGGTTGAGT TTATGCTGCC GGTGCTCAT ACAGTGCGCC TGATTGCGGT	14940
GACTTCTACA TCCGCCAGTC AGGGTGCAAG TTAATTTGAC AGTCAGGTGA ATGGCCGCGA	15000
TTGGCGTGTG GCCTCTGAGT CACCTATTCA ATTAGGGCGA TCACATGGGG GTCATACTTA	15060
ATCAGGCAGG AACCATGTGA CCGAAATTAA AAAAAAAAAA A	15101

(2) INFORMATION FOR SEQ ID NO:15:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 747 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 1..747
- (D) OTHER INFORMATION: /standard_name= "LV ORF 2"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:

ATG CAA TGG GGT CAC TGT GGA GTA AAA TCA GCC AGC TGT TCG TGG ACG	48
Met Gln Trp Gly His Cys Gly Val Lys Ser Ala Ser Cys Ser Trp Thr	
1 5 10 15	
CCT TCA CTG AGT TCC TTG TTA GTG TGG TTG ATA TTG CCA TTT TCC TTG	96
Pro Ser Leu Ser Ser Leu Leu Val Trp Leu Ile Leu Pro Phe Ser Leu	
20 25 30	
CCA TAC TGT TTG GGT TCA CCG TCG CAG GAT GGT TAC TGG TCT TTC TTC	144
Pro Tyr Cys Leu Gly Ser Pro Ser Gln Asp Gly Tyr Trp Ser Phe Phe	
35 40 45	
TCA GAG TGG TTT GCT CCG CGC TTC TCC GTT CGC GCT CTG CCA TTC ACT	192
Ser Glu Trp Phe Ala Pro Arg Phe Ser Val Arg Ala Leu Pro Phe Thr	
50 55 60	
CTC CCG AAC TAT CGA AGG TCC TAT GAA GGC TTG TTG CCC AAC TGC AGA	240
Leu Pro Asn Tyr Arg Arg Ser Tyr Glu Gly Leu Leu Pro Asn Cys Arg	
65 70 75 80	
CCG GAT GTC CCA CAA TTT GCA GTC AAG CAC CCA TTG GGT ATG TTT TGG	288
Pro Asp Val Pro Gln Phe Ala Val Lys His Pro Leu Gly Met Phe Trp	
85 90 95	
CAC ATG CGA GTT TCC CAC TTG ATT GAT GAG ATG GTC TCT CGT CGC ATT	336
His Met Arg Val Ser His Leu Ile Asp Glu Met Val Ser Arg Arg Ile	
100 105 110	
TAC CAG ACC ATG GAA CAT TCA GGT CAA GCG GCC TGG AAG CAG GTG GTT	384
Tyr Gln Thr Met Glu His Ser Gly Gln Ala Ala Trp Lys Gln Val Val	
115 120 125	
GGT GAG GCC ACT CTC ACG AAG CTG TCA GGG CTC GAT ATA GTT ACT CAT	432
Gly Glu Ala Thr Leu Thr Lys Leu Ser Gly Leu Asp Ile Val Thr His	
130 135 140	
TTC CAA CAC CTG GCC GCA GTG GAG GCG GAT TCT TGC CGC TTT CTC AGC	480
Phe Gln His Leu Ala Val Glu Ala Asp Ser Cys Arg Phe Leu Ser	
145 150 155 160	
TCA CGA CTC GTG ATG CTA AAA AAT CTT GCC GTT GGC AAT GTG AGC CTA	528
Ser Arg Leu Val Met Leu Lys Asn Leu Ala Val Gly Asn Val Ser Leu	
165 170 175	
CAG TAC AAC ACC ACG TTG GAC CGC GTT GAG CTC ATC TTC CCC ACG CCA	576
Gln Tyr Asn Thr Thr Leu Asp Arg Val Glu Leu Ile Phe Pro Thr Pro	
180 185 190	
GGT ACG AGG CCC AAG TTG ACC GAT TTC AGA CAA TGG CTC ATC AGT GTG	624
Gly Thr Arg Pro Lys Leu Thr Asp Phe Arg Gln Trp Leu Ile Ser Val	
195 200 205	
CAC GCT TCC ATT TTT TCC TCT GTG GCT TCA TCT GTT ACC TTG TTC ATA	672

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His Ala Ser Ile Phe Ser Ser Val Ala Ser Ser Val Thr Leu Phe Ile
 210 215 220

GTG CTT TGG CTT CGA ATT CCA GCT CTA CGC TAT GTT TTT GGT TTC CAT 720
 Val Leu Trp Leu Arg Ile Pro Ala Leu Arg Tyr Val Phe Gly Phe His
 225 230 235 240

TGG CCC ACG GCA ACA CAT CAT TCG AGC 747
 Trp Pro Thr Ala Thr His His Ser Ser
 245

(2) INFORMATION FOR SEQ ID NO:16:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 249 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:

Met Gln Trp Gly His Cys Gly Val Lys Ser Ala Ser Cys Ser Trp Thr
 1 5 10 15

Pro Ser Leu Ser Ser Leu Leu Val Trp Leu Ile Leu Pro Phe Ser Leu
 20 25 30

Pro Tyr Cys Leu Gly Ser Pro Ser Gln Asp Gly Tyr Trp Ser Phe Phe
 35 40 45

Ser Glu Trp Phe Ala Pro Arg Phe Ser Val Arg Ala Leu Pro Phe Thr
 50 55 60

Leu Pro Asn Tyr Arg Arg Ser Tyr Glu Gly Leu Leu Pro Asn Cys Arg
 65 70 75 80

Pro Asp Val Pro Gln Phe Ala Val Lys His Pro Leu Gly Met Phe Trp
 85 90 95

His Met Arg Val Ser His Leu Ile Asp Glu Met Val Ser Arg Arg Ile
 100 105 110

Tyr Gln Thr Met Glu His Ser Gly Gln Ala Ala Trp Lys Gln Val Val
 115 120 125

Gly Glu Ala Thr Leu Thr Lys Leu Ser Gly Leu Asp Ile Val Thr His
 130 135 140

Phe Gln His Leu Ala Ala Val Glu Ala Asp Ser Cys Arg Phe Leu Ser
 145 150 155 160

Ser Arg Leu Val Met Leu Lys Asn Leu Ala Val Gly Asn Val Ser Leu
 165 170 175

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Gln Tyr Asn Thr Thr Leu Asp Arg Val Glu Leu Ile Phe Pro Thr Pro
 180 185 190

Gly Thr Arg Pro Lys Leu Thr Asp Phe Arg Gln Trp Leu Ile Ser Val
 195 200 205

His Ala Ser Ile Phe Ser Ser Val Ala Ser Ser Val Thr Leu Phe Ile
 210 215 220

Val Leu Trp Leu Arg Ile Pro Ala Leu Arg Tyr Val Phe Gly Phe His
 225 230 235 240

Trp Pro Thr Ala Thr His His Ser Ser
 245

(2) INFORMATION FOR SEQ ID NO:17:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 795 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 1..795
- (D) OTHER INFORMATION: /standard_name= "LV ORF 3"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:

ATG GCT CAT CAG TGT GCA CGC TTC CAT TTT TTC CTC TGT GGC TTC ATC	48
Met Ala His Gln Cys Ala Arg Phe His Phe Phe Leu Cys Gly Phe Ile	
1 5 10 15	
TGT TAC CTT GTT CAT AGT GCT TTG GCT TCG AAT TCC AGC TCT ACG CTA	96
Cys Tyr Leu Val His Ser Ala Leu Ala Ser Asn Ser Ser Ser Thr Leu	
20 25 30	
TGT TTT TGG TTT CCA TTG GCC CAC GGC AAC ACA TCA TTC GAG CTG ACC	144
Cys Phe Trp Phe Pro Leu Ala His Gly Asn Thr Ser Phe Glu Leu Thr	
35 40 45	
ATC AAC TAC ACC ATA TGC ATG CCC TGT TCT ACC AGT CAA GCG GCT CGC	192
Ile Asn Tyr Thr Ile Cys Met Pro Cys Ser Thr Ser Gln Ala Ala Arg	
50 55 60	
CAA AGG CTC GAG CCC GGT CGT AAC ATG TGG TGC AAA ATA GGG CAT GAC	240
Gln Arg Leu Glu Pro Gly Arg Asn Met Trp Cys Lys Ile Gly His Asp	
65 70 75 80	

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AGG TGT GAG GAG CGT GAC CAT GAT GAG TTG TTA ATG TCC ATC CCG TCC	288
Arg Cys Glu Glu Arg Asp His Asp Glu Leu Leu Met Ser Ile Pro Ser	
85 90 95	
GGG TAC GAC AAC CTC AAA CTT GAG GGT TAT TAT GCT TGG CTG GCT TTT	336
Gly Tyr Asp Asn Leu Lys Leu Glu Gly Tyr Tyr Ala Trp Leu Ala Phe	
100 105 110	
TTG TCC TTT TCC TAC GCG GCC CAA TTC CAT CCG GAG TTG TTC GGG ATA	384
Leu Ser Phe Ser Tyr Ala Ala Gln Phe His Pro Glu Leu Phe Gly Ile	
115 120 125	
GGG AAT GTG TCG CGC GTC TTC GTG GAC AAG CGA CAC CAG TTC ATT TGT	432
Gly Asn Val Ser Arg Val Phe Val Asp Lys Arg His Gln Phe Ile Cys	
130 135 140	
GCC GAG CAT GAT GGA CAC AAT TCA ACC GTA TCT ACC GGA CAC AAC ATC	480
Ala Glu His Asp Gly His Asn Ser Thr Val Ser Thr Gly His Asn Ile	
145 150 155 160	
TCC GCA TTA TAT GCG GCA TAT TAC CAC CAC CAA ATA GAC GGG GGC AAT	528
Ser Ala Leu Tyr Ala Ala Tyr Tyr His His Gln Ile Asp Gly Gly Asn	
165 170 175	
TGG TTC CAT TTG GAA TGG CTG CGG CCA CTC TTT TCT TCC TGG CTG GTG	576
Trp Phe His Leu Glu Trp Leu Arg Pro Leu Phe Ser Ser Trp Leu Val	
180 185 190	
CTC AAC ATA TCA TGG TTT CTG AGG CGT TCG CCT GTA AGC CCT GTT TCT	624
Leu Asn Ile Ser Trp Phe Leu Arg Arg Ser Pro Val Ser Pro Val Ser	
195 200 205	
CGA CGC ATC TAT CAG ATA TTG AGA CCA ACA CGA CCG CGG CTG CCG GTT	672
Arg Arg Ile Tyr Gln Ile Leu Arg Pro Thr Arg Pro Arg Leu Pro Val	
210 215 220	
TCA TGG TCC TTC AGG ACA TCA ATT GTT TCC GAC CTC ACG GGG TCT CAG	720
Ser Trp Ser Phe Arg Thr Ser Ile Val Ser Asp Leu Thr Gly Ser Gln	
225 230 235 240	
CAG CGC AAG AGA AAA TTT CCT TCG GAA AGT CGT CCC AAT GTC GTG AAG	768
Gln Arg Lys Arg Lys Phe Pro Ser Glu Ser Arg Pro Asn Val Val Lys	
245 250 255	
CCG TCG GTA CTC CCC AGT ACA TCA CGA	795
Pro Ser Val Leu Pro Ser Thr Ser Arg	
260 265	

(2) INFORMATION FOR SEQ ID NO:18:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 265 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

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(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:

Met Ala His Gln Cys Ala Arg Phe His Phe Phe Leu Cys Gly Phe Ile
 1 5 10 15
 Cys Tyr Leu Val His Ser Ala Leu Ala Ser Asn Ser Ser Ser Thr Leu
 20 25 30
 Cys Phe Trp Phe Pro Leu Ala His Gly Asn Thr Ser Phe Glu Leu Thr
 35 40 45
 Ile Asn Tyr Thr Ile Cys Met Pro Cys Ser Thr Ser Gln Ala Ala Arg
 50 55 60
 Gln Arg Leu Glu Pro Gly Arg Asn Met Trp Cys Lys Ile Gly His Asp
 65 70 75 80
 Arg Cys Glu Glu Arg Asp His Asp Glu Leu Leu Met Ser Ile Pro Ser
 85 90 95
 Gly Tyr Asp Asn Leu Lys Leu Glu Gly Tyr Tyr Ala Trp Leu Ala Phe
 100 105 110
 Leu Ser Phe Ser Tyr Ala Ala Gln Phe His Pro Glu Leu Phe Gly Ile
 115 120 125
 Gly Asn Val Ser Arg Val Phe Val Asp Lys Arg His Gln Phe Ile Cys
 130 135 140
 Ala Glu His Asp Gly His Asn Ser Thr Val Ser Thr Gly His Asn Ile
 145 150 155 160
 Ser Ala Leu Tyr Ala Ala Tyr Tyr His His Gln Ile Asp Gly Gly Asn
 165 170 175
 Trp Phe His Leu Glu Trp Leu Arg Pro Leu Phe Ser Ser Trp Leu Val
 180 185 190
 Leu Asn Ile Ser Trp Phe Leu Arg Arg Ser Pro Val Ser Pro Val Ser
 195 200 205
 Arg Arg Ile Tyr Gln Ile Leu Arg Pro Thr Arg Pro Arg Leu Pro Val
 210 215 220
 Ser Trp Ser Phe Arg Thr Ser Ile Val Ser Asp Leu Thr Gly Ser Gln
 225 230 235 240
 Gln Arg Lys Arg Lys Phe Pro Ser Glu Ser Arg Pro Asn Val Val Lys
 245 250 255
 Pro Ser Val Leu Pro Ser Thr Ser Arg
 260 265

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(2) INFORMATION FOR SEQ ID NO:19:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 549 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 1..549
- (D) OTHER INFORMATION: /standard_name= "LV ORF 4"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:19:

ATG GCT GCG GCC ACT CTT TTC TTC CTG GCT GGT GCT CAA CAT ATC ATG	48
Met Ala Ala Ala Thr Leu Phe Phe Leu Ala Gly Ala Gln His Ile Met	
1 5 10 15	
GTT TCT GAG GCG TTC GCC TGT AAG CCC TGT TTC TCG ACG CAT CTA TCA	96
Val Ser Glu Ala Phe Ala Cys Lys Pro Cys Phe Ser Thr His Leu Ser	
20 25 30	
GAT ATT GAG ACC AAC ACG ACC GCG GCT GCC GGT TTC ATG GTC CTT CAG	144
Asp Ile Glu Thr Asn Thr Thr Ala Ala Ala Gly Phe Met Val Leu Gln	
35 40 45	
GAC ATC AAT TGT TTC CGA CCT CAC GGG GTC TCA GCA GCG CAA GAG AAA	192
Asp Ile Asn Cys Phe Arg Pro His Gly Val Ser Ala Ala Gln Glu Lys	
50 55 60	
ATT TCC TTC GGA AAG TCG TCC CAA TGT CGT GAA GCC GTC GGT ACT CCC	240
Ile Ser Phe Gly Lys Ser Ser Gln Cys Arg Glu Ala Val Gly Thr Pro	
65 70 75 80	
CAG TAC ATC ACG ATA ACG GCT AAC GTG ACC GAC GAA TCA TAC TTG TAC	288
Gln Tyr Ile Thr Ile Thr Ala Asn Val Thr Asp Glu Ser Tyr Leu Tyr	
85 90 95	
AAC GCG GAC CTG CTG ATG CTT TCT GCG TGC CTT TTC TAC GCC TCA GAA	336
Asn Ala Asp Leu Leu Met Leu Ser Ala Cys Leu Phe Tyr Ala Ser Glu	
100 105 110	
ATG AGC GAG AAA GGC TTC AAA GTC ATC TTT GGG AAT GTC TCT GGC GTT	384
Met Ser Glu Lys Gly Phe Lys Val Ile Phe Gly Asn Val Ser Gly Val	
115 120 125	
GTT TCT GCT TGT GTC AAT TTC ACA GAT TAT GTG GCC CAT GTG ACC CAA	432
Val Ser Ala Cys Val Asn Phe Thr Asp Tyr Val Ala His Val Thr Gln	
130 135 140	

CAT ACC CAG CAG CAT CTG GTA ATT GAT CAC ATT CGG TTG CTG CAT	
His Thr Gln Gln His His Leu Val Ile Asp His Ile Arg Leu Leu His	480
145 150 155 160	
TTC CTG ACA CCA TCT GCA ATG AGG TGG GCT ACA ACC ATT GCT TGT TTG	
Phe Leu Thr Pro Ser Ala Met Arg Trp Ala Thr Thr Ile Ala Cys Leu	528
165 170 175	
TTC GCC ATT CTC TTG GCA ATA	
Phe Ala Ile Leu Leu Ala Ile	549
180	

Met	Ala	Ala	Ala	Thr	Leu	Phe	Phe	Leu	Ala	Gly	Ala	Gln	His	Ile	Met
1				5					10					15	
Val	Ser	Glu	Ala	Phe	Ala	Cys	Lys	Pro	Cys	Phe	Ser	Thr	His	Leu	Ser
			20					25					30		
Asp	Ile	Glu	Thr	Asn	Thr	Thr	Ala	Ala	Ala	Gly	Phe	Met	Val	Leu	Gln
		35					40					45			
Asp	Ile	Asn	Cys	Phe	Arg	Pro	His	Gly	Val	Ser	Ala	Ala	Gln	Glu	Lys
	50					55					60				
Ile	Ser	Phe	Gly	Lys	Ser	Ser	Gln	Cys	Arg	Glu	Ala	Val	Gly	Thr	Pro
65					70					75					80
Gln	Tyr	Ile	Thr	Ile	Thr	Ala	Asn	Val	Thr	Asp	Glu	Ser	Tyr	Leu	Tyr
				85					90					95	
Asn	Ala	Asp	Leu	Leu	Met	Leu	Ser	Ala	Cys	Leu	Phe	Tyr	Ala	Ser	Glu
			100					105					110		
Met	Ser	Glu	Lys	Gly	Phe	Lys	Val	Ile	Phe	Gly	Asn	Val	Ser	Gly	Val
		115					120					125			
Val	Ser	Ala	Cys	Val	Asn	Phe	Thr	Asp	Tyr	Val	Ala	His	Val	Thr	Gln
		130				135					140				
His	Thr	Gln	Gln	His	His	Leu	Val	Ile	Asp	His	Ile	Arg	Leu	Leu	His
145					150					155					160

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 603 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: linear

(A) NAME/KEY: CDS
(B) LOCATION: 1..603
(D) OTHER INFORMATION: /standard name= "LV ORF 5"

ATG	AGA	TGT	TCT	CAC	AAA	TTG	GGG	CGT	TTC	TTG	ACT	CCG	CAC	TCT	TGC	48
Met	Arg	Cys	Ser	His	Lys	Leu	Gly	Arg	Phe	Leu	Thr	Pro	His	Ser	Cys	
1				5					10					15		
TTC	TGG	TGG	CTT	TTT	TTG	CTG	TGT	ACC	GGC	TTG	TCC	TGG	TCC	TTT	GCC	96
Phe	Trp	Trp	Leu	Phe	Leu	Leu	Cys	Thr	Gly	Leu	Ser	Trp	Ser	Phe	Ala	
			20					25					30			
GAT	GGC	AAC	GGC	GAC	AGC	TCG	ACA	TAC	CAA	TAC	ATA	TAT	AAC	TTG	ACG	144
Asp	Gly	Asn	Gly	Asp	Ser	Ser	Thr	Tyr	Gln	Tyr	Ile	Tyr	Asn	Leu	Thr	
		35					40					45				
ATA	TGC	GAG	CTG	AAT	GGG	ACC	GAC	TGG	TTG	TCC	AGC	CAT	TTT	GGT	TGG	192
Ile	Cys	Glu	Leu	Asn	Gly	Thr	Asp	Trp	Leu	Ser	Ser	His	Phe	Gly	Trp	
	50					55					60					
GCA	GTC	GAG	ACC	TTT	GTG	CTT	TAC	CCG	GTT	GCC	ACT	CAT	ATC	CTC	TCA	240
Ala	Val	Glu	Thr	Phe	Val	Leu	Tyr	Pro	Val	Ala	Thr	His	Ile	Leu	Ser	
65					70					75					80	
CTG	GGT	TTT	CTC	ACA	ACA	AGC	CAT	TTT	TTT	GAC	GCG	CTC	GGT	CTC	GGC	288
Leu	Gly	Phe	Leu	Thr	Thr	Ser	His	Phe	Phe	Asp	Ala	Leu	Gly	Leu	Gly	
				85					90					95		
GCT	GTA	TCC	ACT	GCA	GGA	TTT	GTT	GGC	GGG	CGG	TAC	GTA	CTC	TGC	AGC	336
Ala	Val	Ser	Thr	Ala	Gly	Phe	Val	Gly	Gly	Arg	Tyr	Val	Leu	Cys	Ser	
				100				105					110			

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GTC TAC GGC GCT TGT GCT TTC GCA GCG TTC GTA TGT TTT GTC ATC CGT	384
Val Tyr Gly Ala Cys Ala Phe Ala Ala Phe Val Cys Phe Val Ile Arg	
115 120 125	
GCT GCT AAA AAT TGC ATG GCC TGC CGC TAT GCC CGT ACC CGG TTT ACC	432
Ala Ala Lys Asn Cys Met Ala Cys Arg Tyr Ala Arg Thr Arg Phe Thr	
130 135 140	
AAC TTC ATT GTG GAC GAC CGG GGG AGA GTT CAT CGA TGG AAG TCT CCA	480
Asn Phe Ile Val Asp Asp Arg Gly Arg Val His Arg Trp Lys Ser Pro	
145 150 155 160	
ATA GTG GTA GAA AAA TTG GGC AAA GCC GAA GTC GAT GGC AAC CTC GTC	528
Ile Val Val Glu Lys Leu Gly Lys Ala Glu Val Asp Gly Asn Leu Val	
165 170 175	
ACC ATC AAA CAT GTC GTC CTC GAA GGG GTT AAA GCT CAA CCC TTG ACG	576
Thr Ile Lys His Val Val Leu Glu Gly Val Lys Ala Gln Pro Leu Thr	
180 185 190	
AGG ACT TCG GCT GAG CAA TGG GAG GCC	603
Arg Thr Ser Ala Glu Gln Trp Glu Ala	
195 200	

(2) INFORMATION FOR SEQ ID NO:22:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 201 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:22:

Met Arg Cys Ser His Lys Leu Gly Arg Phe Leu Thr Pro His Ser Cys	
1 5 10 15	
Phe Trp Trp Leu Phe Leu Leu Cys Thr Gly Leu Ser Trp Ser Phe Ala	
20 25 30	
Asp Gly Asn Gly Asp Ser Ser Thr Tyr Gln Tyr Ile Tyr Asn Leu Thr	
35 40 45	
Ile Cys Glu Leu Asn Gly Thr Asp Trp Leu Ser Ser His Phe Gly Trp	
50 55 60	
Ala Val Glu Thr Phe Val Leu Tyr Pro Val Ala Thr His Ile Leu Ser	
65 70 75 80	
Leu Gly Phe Leu Thr Thr Ser His Phe Phe Asp Ala Leu Gly Leu Gly	
85 90 95	

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Ala Val Ser Thr Ala Gly Phe Val Gly Gly Arg Tyr Val Leu Cys Ser
 100 105 110

Val Tyr Gly Ala Cys Ala Phe Ala Ala Phe Val Cys Phe Val Ile Arg
 115 120 125

Ala Ala Lys Asn Cys Met Ala Cys Arg Tyr Ala Arg Thr Arg Phe Thr
 130 135 140

Asn Phe Ile Val Asp Asp Arg Gly Arg Val His Arg Trp Lys Ser Pro
 145 150 155 160

Ile Val Val Glu Lys Leu Gly Lys Ala Glu Val Asp Gly Asn Leu Val
 165 170 175

Thr Ile Lys His Val Val Leu Glu Gly Val Lys Ala Gln Pro Leu Thr
 180 185 190

Arg Thr Ser Ala Glu Gln Trp Glu Ala
 195 200

(2) INFORMATION FOR SEQ ID NO:23:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 519 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: cDNA

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 1..519
- (D) OTHER INFORMATION: /standard_name= "LV ORF 6"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:23:

ATG GGA GGC CTA GAC GAT TTT TGC AAC GAT CCT ATC GCC GCA CAA AAG	48
Met Gly Gly Leu Asp Asp Phe Cys Asn Asp Pro Ile Ala Ala Gln Lys	
1 5 10 15	
CTC GTG CTA GCC TTT AGC ATC ACA TAC ACA CCT ATA ATG ATA TAC GCC	96
Leu Val Leu Ala Phe Ser Ile Thr Tyr Thr Pro Ile Met Ile Tyr Ala	
20 25 30	
CTT AAG GTG TCA CGC GGC CGA CTC CTG GGG CTG TTG CAC ATC CTA ATA	144
Leu Lys Val Ser Arg Gly Arg Leu Leu Gly Leu Leu His Ile Leu Ile	
35 40 45	
TTT CTG AAC TGT TCC TTT ACA TTC GGA TAC ATG ACA TAT GTG CAT TTT	192
Phe Leu Asn Cys Ser Phe Thr Phe Gly Tyr Met Thr Tyr Val His Phe	
50 55 60	

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CAA TCC ACC AAC CGT GTC GCA CTT ACC CTG GGG GCT GTT GTC GCC CTT	240
Gln Ser Thr Asn Arg Val Ala Leu Thr Leu Gly Ala Val Val Ala Leu	
65 70 75 80	
CTG TGG GGT GTT TAC AGC TTC ACA GAG TCA TGG AAG TTT ATC ACT TCC	288
Leu Trp Gly Val Tyr Ser Phe Thr Glu Ser Trp Lys Phe Ile Thr Ser	
85 90 95	
AGA TGC AGA TTG TGT TGC CTT GGC CGG CGA TAC ATT CTG GCC CCT GCC	336
Arg Cys Arg Leu Cys Cys Leu Gly Arg Arg Tyr Ile Leu Ala Pro Ala	
100 105 110	
CAT CAC GTA GAA AGT GCT GCA GGT CTC CAT TCA ATC TCA GCG TCT GGT	384
His His Val Glu Ser Ala Ala Gly Leu His Ser Ile Ser Ala Ser Gly	
115 120 125	
AAC CGA GCA TAC GCT GTG AGA AAG CCC GGA CTA ACA TCA GTG AAC GGC	432
Asn Arg Ala Tyr Ala Val Arg Lys Pro Gly Leu Thr Ser Val Asn Gly	
130 135 140	
ACT CTA GTA CCA GGA CTT CGG AGC CTC GTG CTG GGC GGC AAA CGA GCT	480
Thr Leu Val Pro Gly Leu Arg Ser Leu Val Leu Gly Gly Lys Arg Ala	
145 150 155 160	
GTT AAA CGA GGA GTG GTT AAC CTC GTC AAG TAT GGC CGG	519
Val Lys Arg Gly Val Val Asn Leu Val Lys Tyr Gly Arg	
165 170	

(2) INFORMATION FOR SEQ ID NO:24:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 173 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:24:

Met Gly Gly Leu Asp Asp Phe Cys Asn Asp Pro Ile Ala Ala Gln Lys	
1 5 10 15	
Leu Val Leu Ala Phe Ser Ile Thr Tyr Thr Pro Ile Met Ile Tyr Ala	
20 25 30	
Leu Lys Val Ser Arg Gly Arg Leu Leu Gly Leu Leu His Ile Leu Ile	
35 40 45	
Phe Leu Asn Cys Ser Phe Thr Phe Gly Tyr Met Thr Tyr Val His Phe	
50 55 60	
Gln Ser Thr Asn Arg Val Ala Leu Thr Leu Gly Ala Val Val Ala Leu	
65 70 75 80	

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(2) INFORMATION FOR SEQ ID NO:25:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 384 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(ix) FEATURE:

(A) NAME/KEY: CDS
(B) LOCATION: 1..384
(D) OTHER INFORMATION: /standard name= "LV ORF 7"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:25:

ATG	GCC	GGT	AAA	AAC	CAG	AGC	CAG	AAG	AAA	AAG	AAA	AGT	ACA	GCT	CCG	48
Met	Ala	Gly	Lys	Asn	Gln	Ser	Gln	Lys	Lys	Lys	Lys	Ser	Thr	Ala	Pro	
1				5				10						15		
ATG	GGG	AAT	GGC	CAG	CCA	GTC	AAT	CAA	CTG	TGC	CAG	TTG	CTG	GGT	GCA	96
Met	Gly	Asn	Gly	Gln	Pro	Val	Asn	Gln	Leu	Cys	Gln	Leu	Leu	Gly	Ala	
			20					25					30			
ATG	ATA	AAG	TCC	CAG	CGC	CAG	CAA	CCT	AGG	GGA	GGA	CAG	GCC	AAA	AAG	144
Met	Ile	Lys	Ser	Gln	Arg	Gln	Gln	Pro	Arg	Gly	Gly	Gln	Ala	Lys	Lys	
		35					40					45				
AAA	AAG	CCT	GAG	AAG	CCA	CAT	TTT	CCC	CTG	GCT	GCT	GAA	GAT	GAC	ATC	192
Lys	Lys	Pro	Glu	Lys	Pro	His	Phe	Pro	Leu	Ala	Ala	Glu	Asp	Asp	Ile	
	50					55				60						

CGG	CAC	CAC	CTC	ACC	CAG	ACT	GAA	CGC	TCC	CTC	TGC	TTG	CAA	TCG	ATC	240
Arg	His	His	Leu	Thr	Gln	Thr	Glu	Arg	Ser	Leu	Cys	Leu	Gln	Ser	Ile	
65					70					75					80	
CAG	ACG	GCT	TTC	AAT	CAA	GGC	GCA	GGA	ACT	GCG	TCG	CTT	TCA	TCC	AGC	288
Gln	Thr	Ala	Phe	Asn	Gln	Gly	Ala	Gly	Thr	Ala	Ser	Leu	Ser	Ser	Ser	
				85					90						95	
GGG	AAG	GTC	AGT	TTT	CAG	GTT	GAG	TTT	ATG	CTG	CCG	GTT	GCT	CAT	ACA	336
Gly	Lys	Val	Ser	Phe	Gln	Val	Glu	Phe	Met	Leu	Pro	Val	Ala	His	Thr	
			100					105					110			
GTG	CGC	CTG	ATT	CGC	GTG	ACT	TCT	ACA	TCC	GCC	AGT	CAG	GGT	GCA	AGT	384
Val	Arg	Leu	Ile	Arg	Val	Thr	Ser	Thr	Ser	Ala	Ser	Gln	Gly	Ala	Ser	
		115					120					125				

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 128 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:26:

Met	Ala	Gly	Lys	Asn	Gln	Ser	Gln	Lys	Lys	Lys	Ser	Thr	Ala	Pro	1	5	10	15	
Met	Gly	Asn	Gly	Gln	Pro	Val	Asn	Gln	Leu	Cys	Gln	Leu	Leu	Gly	Ala	20	25	30	
Met	Ile	Lys	Ser	Gln	Arg	Gln	Gln	Pro	Arg	Gly	Gly	Gln	Ala	Lys	Lys	35	40	45	
Lys	Lys	Pro	Glu	Lys	Pro	His	Phe	Pro	Leu	Ala	Ala	Glu	Asp	Asp	Ile	50	55	60	
Arg	His	His	Leu	Thr	Gln	Thr	Glu	Arg	Ser	Leu	Cys	Leu	Gln	Ser	Ile	65	70	75	80
Gln	Thr	Ala	Phe	Asn	Gln	Gly	Ala	Gly	Thr	Ala	Ser	Leu	Ser	Ser	Ser	85	90	95	
Gly	Lys	Val	Ser	Phe	Gln	Val	Glu	Phe	Met	Leu	Pro	Val	Ala	His	Thr	100	105	110	
Val	Arg	Leu	Ile	Arg	Val	Thr	Ser	Thr	Ser	Ala	Ser	Gln	Gly	Ala	Ser	115	120	125	

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Claims:

1. A purified and isolated nucleic acid comprising a fragmentary portion of the VR-2332 genome between ORF 2 and ORF 7 and having a length sufficient to provide a nucleotide sequence that is unique with respect to the LV virus genome.

2. The nucleic acid as set forth in Claim 1, said portion including a coding region for the expression of a polypeptide capable of inducing an anti-PRRS immune response in swine.

3. The nucleic acid as set forth in Claim 1, including said portion selected from Sequence ID No. 1 and being sufficiently dissimilar from portions of Sequence ID. No. 14 to prevent PCR amplification of portions of said Sequence ID No. 1.

4. The nucleic acid as set forth in Claim 3, including said portion consisting essentially of a sequence selected from a group consisting of Sequence ID Nos. 2, 4, 6, 8, 10, 12, and combinations thereof, together with all complimentary strands and degenerate amino acid residue coding equivalencies that may be obtained by site-directed mutagenesis.

5. The nucleic acid as set forth in Claim 3, including said portion consisting essentially of a sequence selected from a group consisting of Sequence ID Nos. 2, 4, 6, 8, 10, and 12, as well as inverse complimentary sequences depending from Sequence ID Nos. 2, 4, 6, 8, 10, and 12.

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6. The nucleic acid as set forth in Claim 5, said group consisting of Sequence ID No. 1 from positions 2783 to 2801, the inverse compliment of Sequence ID No. 1 from positions 3271 to 3289, Sequence ID No. 1 from positions 2289 to 2307, the inverse compliment of Sequence ID No. 1 from positions 2862 to 2880, Sequence ID No. 14 from positions 14112 to 14131, the inverse compliment of Sequence ID No. 14 from positions 14551 to 14570, Sequence ID No. 14 from positions 14575 to 14594, and the inverse compliment of Sequence ID NO. 14 from positions 14955 to 14974, sequence ID No. 1 from positions 2814 to 2832, the inverse compliment of Sequence ID No. 1 from positions 3273 to 3291, Sequence ID No. 1 from positions 2816 to 2834, and the inverse compliment of Sequence ID No. 1 from positions 3181 to 3198.

7. A chimeric vector for use in expressing viral proteins from a host cell, comprising a promoter and a termination sequence connected by a coding region insert including a fragmentary portion of the VR-2332 genome between ORF 2 and ORF 7, said insert having a length sufficient to provide a nucleotide sequence unique with respect to the LV virus genome, together with all degenerate amino acid residue coding equivalencies that may be obtained by site-directed mutagenesis.

8. The vector as set forth in Claim 7, including said insert consisting essentially of a sequence selected from a group consisting of Sequence ID Nos. 2, 4, 6, 8, 10, and 12, as well as inverse complimentary sequences depending from Sequence ID Nos. 2, 4, 6, 8, 10, and 12.

9. A vaccine for immunizing animals against a VR-2332 form of PRRS, comprising a polypeptide-coding region replicating a nucleotide sequence selected as a portion of Sequence ID No. 1 and having a length sufficient to provide a nucleotide sequence unique in comparison with respect to Sequence ID No. 14.

10. The vaccine as set forth in Claim 9, said coding region consisting essentially of a member selected from the group consisting of Sequence ID Nos. 2, 4, 6, 8, 10, and 12, and combinations thereof.

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11. A vaccine for immunizing animals against a VR-2332 form of PRRS, comprising a VR-2332 amino acid residue sequence having a length sufficient to provide uniqueness in comparison with respect to LV virus amino acid residue sequences.

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12. The vaccine as set forth in Claim 11, said VR-2332 amino acid residue sequence consisting essentially of a sequence selected from the group consisting of Sequence ID Nos. 3, 5, 7, 9, 11, and 13, and combinations thereof.

10

13. A diagnostic assay for distinguishing between PRRS-causative viral strains, said assay comprising the steps of:

providing PCR oligonucleotide primers capable of selectively amplifying fragmentary genomic portions of a wild-type PRRS-causative virus;

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obtaining a sample including cDNA derived from swine exhibiting PRRS clinical signs; and

using said primers in a polymerase chain reaction under conditions capable of selective amplification of cDNA from said PRRS-causative virus in said sample.

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14. The assay as set forth in Claim 13, said PRRS-causative virus being selected from a group consisting of VR-2332 and LV virus.

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15. The assay as set forth in Claim 14, including said primers being selected from a group consisting of fragmentary portions of Sequence ID No. 1, complimentary fragments of Sequence ID No. 1, fragmentary portions of Sequence ID No. 14, and complimentary fragments of Sequence ID No. 14, said primers being unique in comparison with respect to Sequence ID No. 14 when said primers derive from Sequence ID No. 1, said primers being unique in comparison with respect to Sequence ID No. 1 when said primers derive from Sequence ID No. 14;

30

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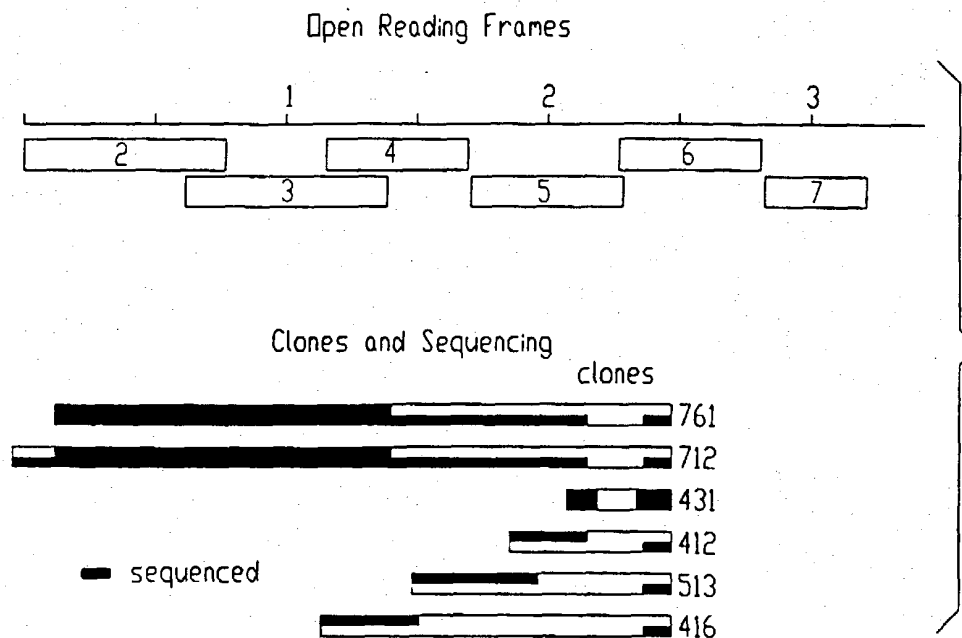
16. A method of vaccinating an animal against VR-2332-caused PRRS, said method comprising the steps of:

providing a vaccine including at least one material selected from a group consisting of VR-2332 based polypeptides and VR-2332 based nucleic acids; and

administering said vaccine to said animal in a manner permitting said animal to develop an immune response to said material.

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1/10

*Fig. 1.*

V E F S L P T H H T V R L I R V T A S P
 3121 TGGAGTTTAGTTTGCCTACGCATCACTGTGCGCCTGATCCGCGTCACAGCATCACCT
 S A *
 3181 CAGCATGATGGCTGGCATTCTTGAGGCATCTCAGTGTGTTGAATTGGAAGAATGTGTGGT
 3241 GAATGGCACTGATTGACATTGTGCCTCTAAGTCACCTATTCAATTAGGGCGACCGTGTGG
 3301 GGGTGAGATTTAATTGGCGAGAACCATGCGGCCGAAATTAATAAAAAAAAAAAAAAAAAA

Fig. 2D.

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Fig. 2A.

DRF 2 M K W G P C K A F L T K L A N F L W M L
1 ATGAAATGGGGTCCATGCAAAGCCTTTTGGACAAAATTGGCCAACTTTTGTGGATGCTT
 S R S S W C P L L I S L Y F W P F C L A
61 TCACGGAGTTCTTGGTGTCCATTGTTGATATCATTATATTTTGGCCATTTTGTGGCT
 S P S P V G W W S F A S D W F A P R Y S
121 TCACCATCGCCGGTGGCTGGTGGTCTTTTGCATCAGATTGGTTTGTCCGCGATACTCC
 V R A L P F T L S N Y R R S Y E A F L S
181 GTACGCGCCCTGCCATTCACTCTGAGCAATTACAGAAGATCTTATGAGGCCTTTCTTTCC
 Q C Q V D I P T W G T K H P L G M L W H
241 CAGTGCCAAGTGGACATTCCCACCTGGGGAATAACATCCCTTTGGGATGCTTTGGCAC
 H K V S T L I D E M V S R R M Y R I M E
301 CATAAGGTGTCAACCCTGATTGATGAAATGGTGTGCGTGAATGTACCGCATCATGGAA
 K A G Q A A W K Q V V S E A T L S R I S
361 AAAGCAGGGCAGGCTGCCTGGAAACAGGTGGTGAGCGAGGCTACGCTGTCTCGCATTAGT
 S L D V V A H F Q H L A A I E A E T C K
421 AGTTTGGATGTGGTGGCTCATTTTCAGCATCTAGCCGCCATTGAAGCCGAGACCTGTAAA
 Y L A S R L P M L H N L R M T G S N V T
481 TATTTGGCCTCCCGGCTGCCCATGCTACACAACCTGCGCATGACAGGGTCAAATGTAACC
 I V Y N S T L N Q V F A I F P T P G S R
541 ATAGTGTATAATAGCACTTTGAATCAGGTGTTTGCTATTTTCCAACCCCTGGTTCCCGG

DRF 3 M V N S C T F L H I F L
 P K L H D F Q Q W L I A V H S S I F S S
601 CCAAAGCTTCATGATTTTCAGCAATGGTTAATAGCTGTACATTCTCCATATTTCTCT
 C C S F L Y S F C C A V V A G S N T T Y
 V A A S C T L F V V L W L R V P I L R T
661 GTTGCAGCTTCTTGTACTCTTTTGTGTGCTGTGGTTGCGGGTTCCAATACTACGTACT
 C F W F P L V R G N F S F E L T V N Y T
 V F G F R W L G A I F L S N S Q *
721 GTTTTTGGTTTCCGCTGGTTAGGGGCAATTTTCTTTGAACTCACAGTGAATTACACGG
 V C P P C L T R Q A A T E I Y E P G R S
781 TGTGTCCACCTTGCCCTACCCGGCAAGCAGCCACAGAGATCTACGAACCCGGTAGGTCTC
 L W C R I G Y D R C G E D D H D E L G F
841 TTTGGTGCAGGATAGGGTATGACCGATGTGGGGAGGACGATCATGACGAGCTAGGGTTTA
 M I P P G L S S E G H L T G V Y A W L A
901 TGATACCGCTGGCCTCTCCAGCGAAGGCCACTTGACTGGTGTGTTACGCCTGGTTGGCGT
 F L S F S Y T A Q F H P E I F G I G N V
961 TCTTGCTTCAGCTACACGGCCAGTTCCATCCCGAGATATTCGGGATAGGGAATGTGA
 S R V Y V D I K H Q L I C A E H D G Q N
1021 GTCGAGTTTATGTTGACATCAAACATCAACTCATCTGCGCCGAACATGACGGGCAGAACA

Fig. 2B.

T T L P R H D N I S A V F Q T Y Y Q H Q
 1081 CCACCTTGCTCGTCATGACAACATTTACGCCGTGTTTCAGACCTATTACCAACATCAAG
 DRF 4 M A S S L L F L V V G
 V D G G N W F H L E W L R P F F S S W L
 1141 TCGACGGCGGCAATTGGTTTACCTAGAAATGGCTTCGTCCCTTCTTTTCTCGTGGTTGG
 F K C L L V S Q A F A C K P C F S S S L
 V L N V S W F L R R S P A N H V S V R V
 1201 TTTTAAATGTCTCTTGGTTTCTCAGGCGTTCCGCTGCAAACCATGTTTCAGTTCGAGTCT
 A D I K T N T T A A A S F A V L Q D I S
 L Q I L - R P T P P Q R Q A L L S S K T S
 1261 TGCAGATATTAAGACCAACACCACCGCAGCGGCAAGCTTTGCTGTCCTCCAAGACATCAG
 C L R H R D S A S E A I R K I P Q C R T
 V A L G I A T R P L R R F A K S L S A V
 1321 TTGCCTTAGGCATCGCGACTCGGCTCTGAGGCGATTGCAAAATCCCTCAGTGGCGTAC
 A I G T P V Y V T I T A N V T D E N Y L
 R R *
 1381 GGCGATAGGGACACCCGTGTATGTTACCATCACAGCCAATGTGACAGATGAGAATTATTT
 H S S D L L M L S S C L F Y A S E M S E
 1441 ACATTCTTCTGATCTCCTCATGCTTTCTTCTTGCCCTTTCTATGCTTCTGAGATGAGTGA
 K G F K V V F G N V S G I V A V C V N F
 1501 AAAGGGATTTAAGGTGGTATTTGGCAATGTGTCAGGCATCGTGGCTGTGTGTGCAATTT
 T S Y V Q H V K E F T Q R S L V V D H V
 1561 TACCAGCTACGTCCAACATGTCAAGGAGTTTACCCAACGCTCCCTGGTGGTGCACCATGT
 R L L H F M T P E T M R W A T V L A C L
 1621 GCGGTTGCTCCATTTTCATGACACCTGAGACCATGAGGTGGGCAACTGTTTTAGCCTGTCT
 DRF 5 F A I L L A I * M L E K C L T A
 1681 TTTTGCCATTCTGTTGGCAATTTGAATGTTAAGTATGTTGGAGAAATGCTTGACCGCGG
 G C C S R L L S L W C I V P F C F A V L
 1741 GCTGTTGCTCGCGATTGCTTTCTTTGTGGTGTATCGTGCCGTTCTGTTTTGCTGTGCTCG
 A N A S N D S S S H L Q L I Y N L T L C
 1801 CCAACGCCAGCAACGACAGCAGCTCCCATCTACAGCTGATTTACAACCTGACGCTATGTG
 E L N G T D W L A N K F D W A V E S F V
 1861 AGCTGAATGGCACAGATTGGCTAGCTAACAATTTGATTGGGAGTGGAGAGTTTTGTCA
 I F P V L T H I V S Y G A L T T S H F L
 1921 TCTTTCCCGTTTTGACTCACATTGTCTCCTATGGTGCCCTCACTACCAGCCATTTCTTG
 D T V A L V T V S T A G F V H G R Y V L
 1981 ACACAGTCGCTTTAGTCACTGTGTCTACCGCCGGGTTGTTACGGGCGGTATGTCCTAA
 S S I Y A V C A L A A L T C F V I R F A
 2041 GTAGCATCTACGCGTCTGTGCCCTGGCTGCGTTGACTTGCTTCGTCATTAGGTTTGCAA

Fig. 2C.

2101 K N C M S W R Y A C T R Y T N F L L D T
AGAATTGCATGTCCTGGCGCTACGCGTGTACCAGATATACCAACTTTCTTCTGGACACTA
2116 K G R L Y R W R S P V I I E K R G K V E
AGGGCAGACTCTATCGTTGGCGGTGCGCTGTCATCATAGAGAAAAGGGGCAAAGTTGAGG
V E G H L I D L K R V V L D G S V A T P
2221 TCGAAGGTCATCTGATCGACCTCAAAGAGTTGTGCTTGATGGTCCGTGGCAACCCCTA
ORF 6 M G S S L D D F C H D S T
I T R V S A E Q W G R P *
2281 TAACCAGAGTTTCAGCGGAACAATGGGGTGGTCTTAGATGACTTCTGTCATGATAGCAC
A P Q K V L L A F S I T Y T P V M I Y A
2341 GGCTCCACAAAAGGTGCTTTTGGCGTTTTCTATTACCTACACGCCAGTGATGATATATGC
L K V S R G R L L G L L H L L I F L N C
2401 CCTAAAGGTGAGTCGCGGCCGACTGCTAGGGCTTCTGCACCTTTTGATCTTCCTGAATTG
A F T F G Y M T F A H F Q S T N K V A L
2461 TGCTTTCACCTTCGGGTACATGACTTTCGCGCACTTCAGAGTACAAATAAGGTGCGGCT
T M G A V V A L L W G V Y S A I E T W K
2521 CACTATGGGAGCAGTAGTTGCACTCCTTTGGGGGTGTACTCAGCCATAGAAACCTGGAA
F I T S R C R L C L L G R K Y I L A P A
2581 ATTCATCACCTCCAGATGCCGTTTGTGCTTGCTAGGCCGCAAGTACATTCTGGCCCCCTGC
H H V E S A A R F H P I A A N D N H A F
2641 CCACCACGTTGAAAGTGCCGCACGGTTTCATCCGATTGCGGCAAATGATAACCACGCATT
V V R R P G S T T V N G T L V P G L K S
2701 TGTCGTCCGGGTCCCGGCTCCACTACGGTCAACGGCACATTGGTGCCCGGGTTAAAAAG
ORF 7 M
L V L G G R K A V K Q G V V N L V K Y A
2761 CCTCGTGTGGGTGGCAGAAAAGCTGTTAAACAGGGAGTGGTAAACCTTGTCAAATATGC
P N N N G K Q T E E K K G D G Q P V N Q
K *
2821 CAAATAACAACGGCAAGCAGACAGAAGAGAAGAAGGGGGATGGCCAGCCAGTCAATCAGC
L C Q M L G K I I A Q Q N Q S R G K G P
2881 TGTGCCAGATGCTGGTAAGATCATCGCTCAGCAAAACCAGTCCAGAGGCAAGGGACCGG
G K K N K K K N P E K P H F P L A T E D
2941 GAAAGAAAAATAAGAAGAAAAACCCGGAGAAGCCCCATTTCTCTAGCGACTGAAGATG
D V R H H F T P S E R Q L C L S S I Q T
3001 ATGTCAGACATCACTTTACCCCTAGTGAGCGGCAATTGTGTCTGTCTGTCATCCAGACCG
A F N Q G A G T C T L S D S G R I S Y T
3061 CCTTTAATCAAGGCGCTGGGACTTGCACCCTGTCAGATTTCAGGGAGGATAAGTTACACTG

Fig. 5A.

2332 1 Mlekcltagccsrllslwciwpfc-favlanA-sNdssSh QIIYNLTICELNGTDMlankFdwAVESFVif
 LV 1 MrcshklgrfltpshcfwllflCtglswsfAdgNgdSSstyQyIYNLTICELNGTDMlsshfgwAVEtfVly

71 PVITHlvSyGaLITSHFIDtvaLvtVSTAGFVhGRYVLSSiYavCAIAAItCFVIRfAKNCMSwRYAcTRYt
 73 PvaTHIISIGFLITSHFFDalgLgaVSTAGFVgGRYVLCsvYgaCAFAAFvCFVIRaAKNCMacRYArTRfT

143 NFIIIDtkGRlyRWvSPviiiEKrGKvEveGhLi dIKrVVLdGsvAtP:TRvSAEQWgrp
 145 NFivDdrGRvhrWkSPivvEKIGKaEVdGnLvtiKhVVLeGvkaAqPI TRtSAEQwea

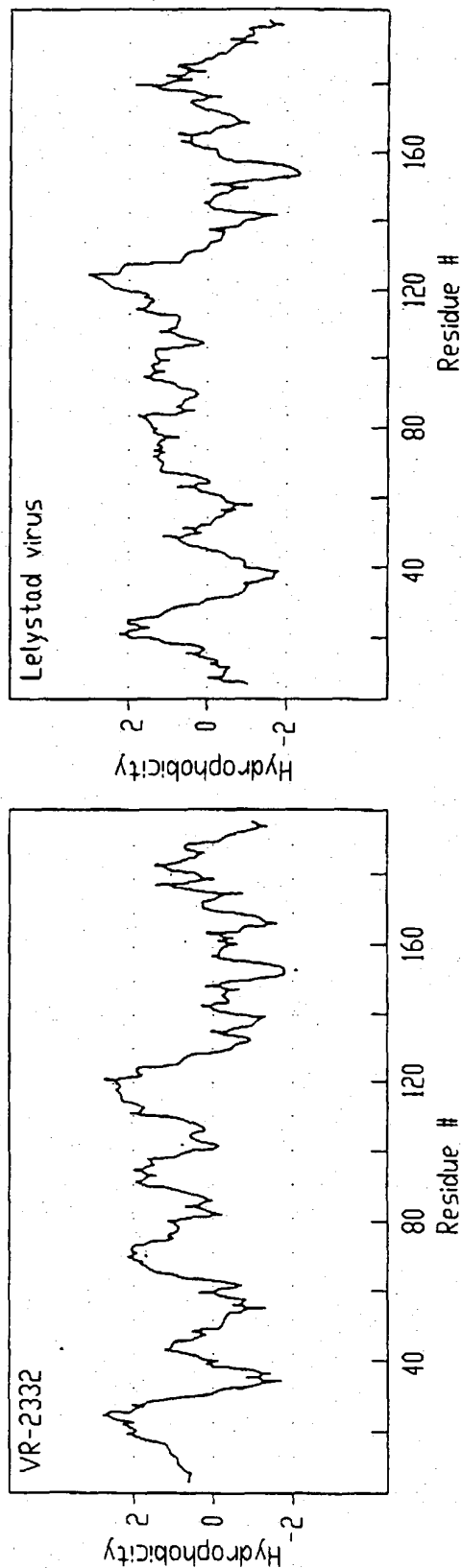


Fig. 5B.

Fig. 5C.

Fig. 6A.

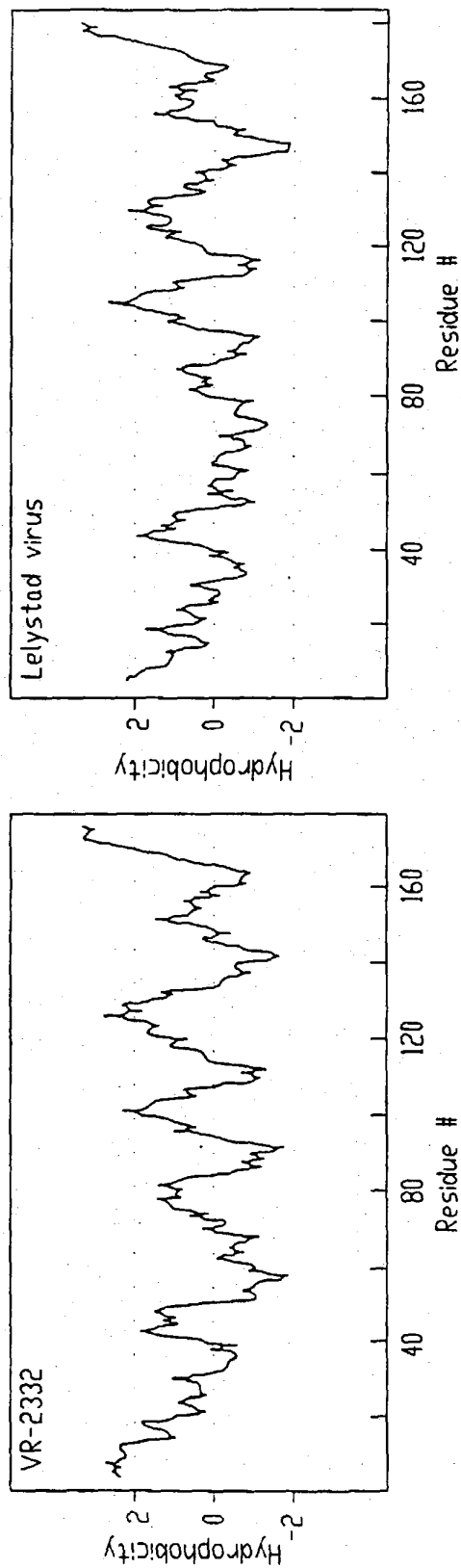
[illegible]

Fig. 6B.

Fig. 6C.

Fig. 7A.

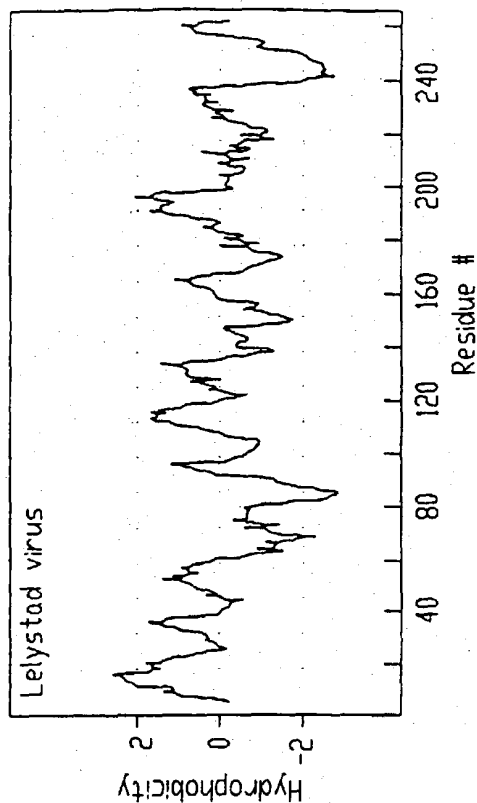
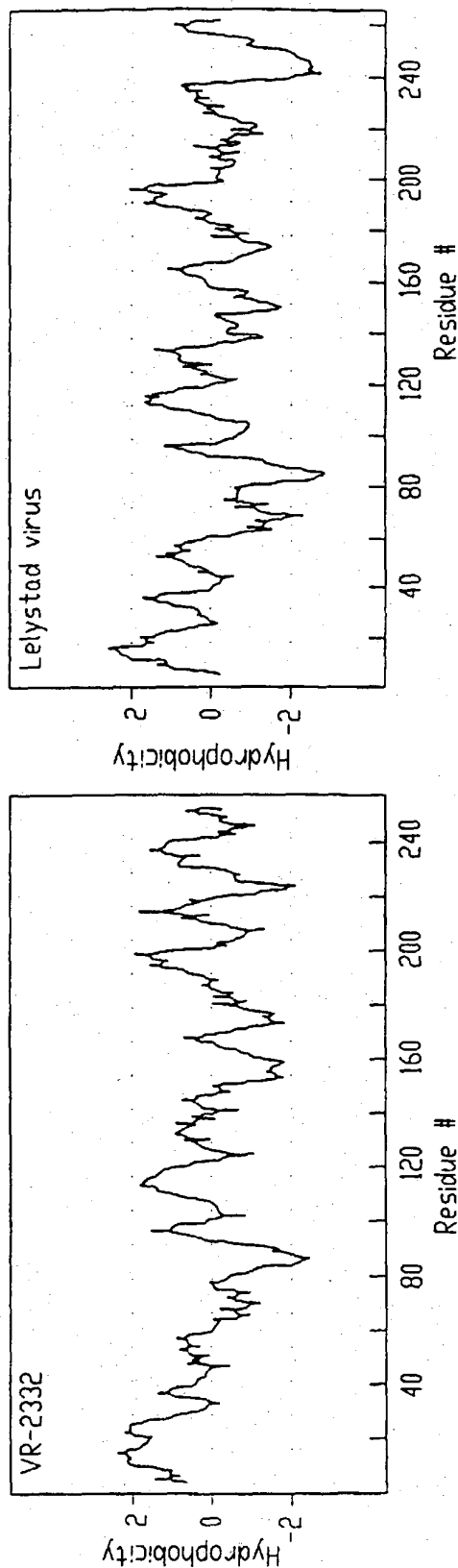
[illegible]

Fig. 8A.

2332 1 MkwGpCkaFltKIAnFlWmLSrSSwcpILVslylwPFCLoSPSpGwSfaSdMfAPRySVRALPFTILsNYR
 LV 1 MqwGhC---gvKsAscsWtpSISStlvwLlIpfsIPyCLgSPSqdGyWSFFSewFAPRfSVRALPFTILpNYR
 73 RSYEaFLsqQqvDiPtwgTKHPLGMLVHhkVStLIDEMVSRRmYr;MEkaGQAAWKQVSEATLsr;SsLDv
 70 RSYEGtlpnCrpDvPqfavKHPLGMFMHmrVShLIDEMVSRR;YqtmEhSGQAAWKQVAgEATLtkISgLDi
 145 VoHFQHLAAIEAetCkylLoSRLpMLhNLrmtGsnVt;vYNsTLnqVfaIFPTPGSRPKLhDFqQWLlAvHsS
 142 VtHFQHLAAVEAdSCrfLSsRLvMLkNL-avG-NVsIqYntTLdrVelIFPTPGtRPKLtDFrQWLIsVHaS
 217 IFSSVaoScTLFvVLWLRvPiLRtVFGFrMgAIFISnSg
 212 IFSSVAsvTLF:VLWLRiPaLRyVFGfhwptAthhS-S

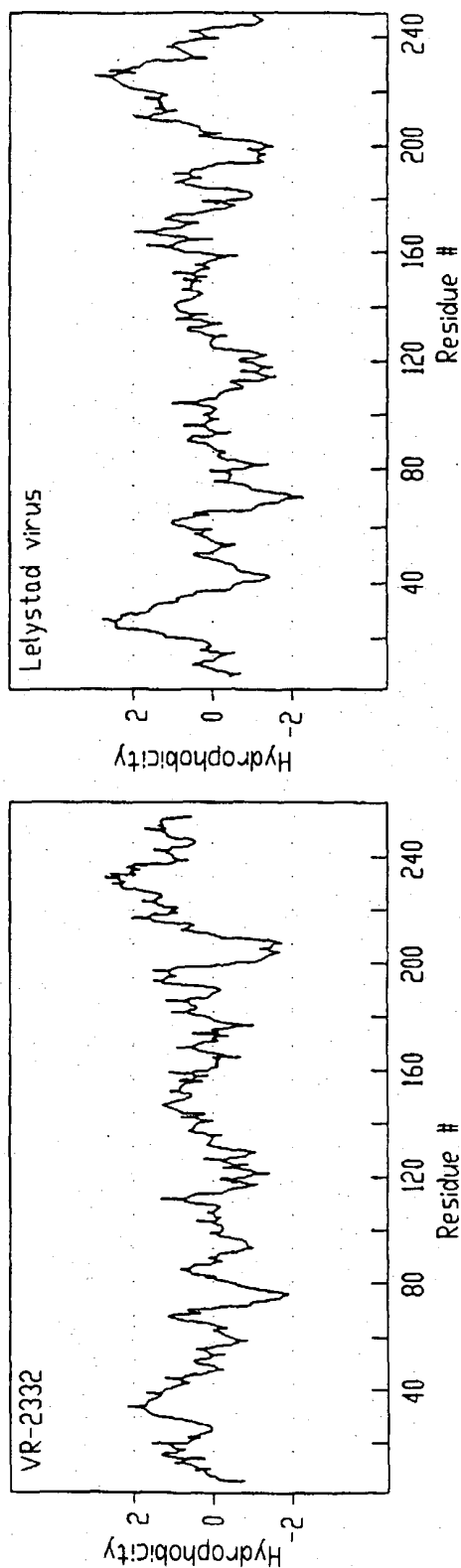


Fig. 8B.

Fig. 8C.

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VR-2332	1	TGGCTGGCATTCTTGAGGCATCTCAGTGTTGAATTGGAAGAATGTGTGGTGAAATGGCAC
LV	1	 AATTGACAGTCAGGTGAATGGCCG
VR-2332	62	TGATTGACATTGTGCCCTCTAAGTCACCTATTCAATTAGGGCGACCGTGTGGGGTGAGATT
LV	25	 CGATTGGCGTGTGGCCCTCTGAGTCACCTATTCAATTAGGGCGATCACA TGGGGTCATACT
VR-2332	123	TAAT TGGCGAGAACCATGCGCCGAAATTAAAAAATAAAAAAAAAAAAAA
LV	86	 TAATCAGGCAGGAACCATGTGACCGAAATTAAAAAATAAAAAAAAAAAAAA

Fig. 9.

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US95/09927

A. CLASSIFICATION OF SUBJECT MATTER IPC(6) : A61K 39/12; C12N 15/14, 15/63 US CL : 424/186.1, 218.1, 815; 435/320; 536/23.72, 24.3 According to International Patent Classification (IPC) or to both national classification and IPC				
B. FIELDS SEARCHED Minimum documentation searched (classification system followed by classification symbols) U.S. : 424/186.1, 218.1, 815; 435/320; 536/23.72, 24.3 Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched Electronic data base consulted during the international search (name of data base and, where practicable, search terms used) Please See Extra Sheet.				
C. DOCUMENTS CONSIDERED TO BE RELEVANT				
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.		
X	WO 93/03760 (COLLINS ET AL.) 04 MARCH 1993, see page 3 and claims.	9, 11, 16		
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Y		1-8, 10, 12-15		
Y	Journal of Veterinary Diagnostic Investigation, Volume 4, Number 2, issued April 1992, Benfield et al., "Characterization of swine infertility and respiratory syndrome (SIRS) virus (isolate ATCC VR-2332)", pages 127-133, see entire document.	1-16		
<input checked="" type="checkbox"/> Further documents are listed in the continuation of Box C. <input type="checkbox"/> See patent family annex.				
<table border="0"> <tr> <td> * Special categories of cited documents: "A" document defining the general state of the art which is not considered to be of particular relevance "E" earlier document published on or after the international filing date "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified) "O" document referring to an oral disclosure, use, exhibition or other means "P" document published prior to the international filing date but later than the priority date claimed </td> <td> "T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention "X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone "Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art "Z" document member of the same patent family </td> </tr> </table>			* Special categories of cited documents: "A" document defining the general state of the art which is not considered to be of particular relevance "E" earlier document published on or after the international filing date "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified) "O" document referring to an oral disclosure, use, exhibition or other means "P" document published prior to the international filing date but later than the priority date claimed	"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention "X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone "Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art "Z" document member of the same patent family
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Date of the actual completion of the international search 29 SEPTEMBER 1995		Date of mailing of the international search report 19 OCT 1995		
Name and mailing address of the ISA/US Commissioner of Patents and Trademarks Box PCT Washington, D.C. 20231 Facsimile No. (703) 305-3230		Authorized officer <i>R. Freese</i> LAWRENCE J. CARROLL, II Telephone No. (703) 308-0196		

INTERNATIONAL SEARCH REPORT

Int. application No.
PCT/US95/09927

C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT		
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	Archives of Virology, Volume 135, issued 1994, Suarez et al., "Direct detection of the porcine reproductive and respiratory syndrome (PRRS) virus by reverse polymerase chain reaction (RT-PCR)", pages 89-99, see summary and page 97 first full paragraph.	1-6, 13-16
Y	Journal of General Virology, Volume 75, Number 7, issued July 1994, Meng et al., "Molecular cloning and nucleotide sequencing of the 3'-terminal genomic RNA of the porcine reproductive and respiratory syndrome virus", pages 1795-1801, see entire document.	1-6
Y	Journal of General Virology, Volume 75, Number 3, issued March 1994, Mardassi et al., "Identification of the major differences in the nucleocapsid protein genes of a Quebec strain and European strains of porcine reproductive and respiratory syndrome virus", pages 681-685, see figure 2.	1-6
P, Y	Veterinary Microbiology, Volume 44, issued 1995, Katz et al., "Antigenic differences between European and American isolates of porcine reproductive and respiratory syndrome virus (PRRSV) are encoded by the carboxyterminal portion of viral open reading frame 3", pages 65-76, see entire document.	1-16

INTERNATIONAL SEARCH REPORT

International application No.

PCT/US95/09927

B. FIELDS SEARCHED

Electronic data bases consulted (Name of data base and where practicable terms used):

APS, DIALOG, MEDLINE, INPADOC, DERWENT WPI, AGRICOLA, CABA
SEARCH TERMS: PORCINE REPRODUCTIVE AND RESPIRATORY SYNDROME, PRRS, SWINE
RESPIRATORY AND INFERTILITY SYNDROME, SIRS, PORCINE EPIDEMIC ABORTION AND RESPIRATORY
SYNDROME, PEARLS, VR 2332